Higher Sclerostin/SOST expression is associated with lower percentage of circulatory blasts and better prognosis in patients with myelofibrosis

Lucijanić, Marko; Livun, Ana; Tupek, Katarina Marija; Štoos-Veić, Tajana; Pejša, Vlatko; Kušec, Rajko

Source / Izvornik: Annals of Hematology, 2018, 97, 1293 - 1294

Journal article, Published version Rad u časopisu, Objavljena verzija rada (izdavačev PDF)

https://doi.org/10.1007/s00277-018-3294-9

Permanent link / Trajna poveznica: https://urn.nsk.hr/urn:nbn:hr:105:406711

Rights / Prava: In copyright/Zaštićeno autorskim pravom.

Download date / Datum preuzimanja: 2025-02-16



Repository / Repozitorij:

<u>Dr Med - University of Zagreb School of Medicine</u> Digital Repository





Središnja medicinska knjižnica

Lucijanić M., Livun A., Tupek K. M., Štoos-Veić T., Pejša V., Kušec R. (2018) *Higher Sclerostin/SOST expression is associated with lower percentage of circulatory blasts and better prognosis in patients with myelofibrosis.* Annals of Hematology, 97 (7). pp. 1293-1294. ISSN 0939-5555

http://www.springer.com/journal/277

http://link.springer.com/journal/277

The final publication is available at Springer via https://doi.org/10.1007/s00277-018-3294-9

http://medlib.mef.hr/3404

University of Zagreb Medical School Repository http://medlib.mef.hr/ Title: Higher Sclerostin/SOST expression is associated with lower percentage of circulatory blasts and

better prognosis in patients with myelofibrosis

Authors: Marko Lucijanic¹, Ana Livun², Katarina Marija Tupek², Tajana Stoos-Veic^{3,4}, Vlatko

Pejsa^{1,5}, Rajko Kusec^{1,2,5}

Affiliations:

¹ Hematology Department, University Hospital Dubrava, Zagreb, Croatia

² Division of Molecular Diagnosis and Genetics, Clinical Department of Laboratory Diagnostics,

University Hospital Dubrava, Zagreb, Croatia

³ Department of Clinical Cytology and Cytometry, University Hospital Dubrava, Zagreb, Croatia

⁴ University of Osijek, Faculty of Medicine, Osijek, Croatia

⁵ University of Zagreb, School of Medicine, Zagreb, Croatia

Corresponding author: Marko Lucijanic, MD PhD, Hematology Department, University Hospital

Dubrava, Av. Gojka Suska 6, 10000 Zagreb. Email: markolucijanic@yahoo.com

ORCID: http://orcid.org/0000-0002-1372-2040

1

Dear Editor,

Sclerostin (a product of *SOST* gene) acts as a main negative regulator of bone metabolism, exerting its properties through inhibition of canonical-WNT signaling-pathway (cWNT) in osteoblasts [1]. It is produced by osteocytes and bone marrow (BM) cells. cWNT activation is implicated in pathogenesis of Philadelphia chromosome negative myeloproliferative neoplasms (Ph- MPNs) [2,3], diseases characterized by remodelling of BM stroma and development of BM fibrosis/osteosclerosis during course of the disease. We aimed to investigate Sclerostin/*SOST* expression in BM tissues of patients with primary (PMF) and secondary post Ph- MPN myelofibrosis (SMF) and to assess its clinical correlations.

We retrospectively investigated Sclerostin/SOST expression in BM of 66 diseased patients (51 PMF, 15 post-PV/post-ET-SMF, diagnoses were established according to the WHO [4] and the IWG-MRT [5] criteria) and 18 age- and sex-matched controls (limited-stage aggressive Non-Hodgkin-lymphoma patients without BM involvement) using immunohistochemistry (IHC; Sclerostin 21933-1-AP rabbit polyclonal Proteintech primary antibody) and real-time polymerase-chain-reaction (RT-PCR; SOST Hs00228830_ml Thermo Fischer Scientific TaqMan assay; evaluated from BM aspirates). Sclerostin expression was expressed as a percentage of positive cells. SOST mRNA expression was normalized to Abl and expressed as a ΔCT value. Correlations with clinical parameters were made. Optimal cut-off values for survival were determined using the ROC-curve-analysis. The Mann-Whitney-U-test, the Spearman-rank-correlation, the Cox-Mantel-log-rank-test [6] and the Cox-regression-analysis were used. Analyses were done using MedCalc-Statistical-Software ver.18 (MedCalc Software bvba, Ostend, Belgium). P values <0.05 were considered statistically significant. All procedures were in accordance with the ethical standards of the responsible committee on human experimentation (institutional and national) and with the Helsinki Declaration of 1975, as revised in 2008. All patients provided written informed consent.

Median age of patients was 67 years, 59% were males, 67% had grade II-III BM fibrosis, 70%, 11% and 2% were *JAK2*, *CALR* and *MPL* mutated, respectively. Median follow up of our cohort was 74

months with median overall survival of 69 months. Sclerostin/SOST expression did not significantly differ between PMF and SMF, nor between patients and controls. However, non-significantly higher expression was observed in myelofibrosis patients (both PMF and SMF) than in controls, result was near statistical significance for SOST (P=0.056). In diseased patients, higher Sclerostin expression measured by IHC was significantly correlated with lower percentage of circulatory blasts (Rho -0.28, P=0.042) and transfusion dependency (P=0.049). Higher SOST expression measured by RT-PCR was similarly significantly correlated with lower percentage of circulatory blasts (Rho -0.44, P=0.042), but also higher platelets (Rho 0.4, P=0.031) and smaller spleen size (Rho -0.6, P=0.001). We found no significant association of Sclerostin/SOST expression with JAK2, CALR and MPL mutation status or degree of bone marrow fibrosis. Patients with higher Sclerostin expression (HR=0.35, P=0.006) and higher SOST expression (HR=0.44, P=0.044) had superior overall survival than patients presenting with lower Sclerostin/SOST expression as shown in a Figure. This association remained significant for SOST (HR=0.21, P=0.025) after adjusting for age, gender and circulatory blasts (HR=1.06, P=0.002). Several interesting observations emerge from our study. First, Sclerostin expression might affect stem-cell-mobilization. SOST deficient mice were shown to have reduced expression of CXCL12 [7] which is important for this process, and reverse situation with higher Sclerostin/SOST expression might be possible. Additionally, patients with higher Sclerostin/SOST expression experienced improved survival, effect which might be prognostically independent of reduction in circulatory blasts (known prognostic parameter in myelofibrosis [8]) and which is probably mediated through cWNT inhibition. Due to the limited number of patients, our study was probably underpowered to detect statistical significance of some existing clinical correlations. Nevertheless, our findings emphasize the role of bone metabolism regulating cytokines, such as Sclerostin, in pathogenesis of Ph- MPNs and suggest that cWNT inhibition might be an interesting therapeutic approach in myelofibrosis patients.

Acknowledgements: The study was funded by University of Zagreb Research grant BM068 project

1101439 to RK

Conflict of interest: The authors report no conflicts of interest.

Informed consent: All subjects provided written informed consent.

References:

1. Shahnazari M, Yao W, Corr M, Lane NE (2008) Targeting the Wnt Signaling Pathway to Augment

Bone Formation. Current osteoporosis reports 6 (4):142-148

2. Lucijanic M, Livun A, Tomasovic-Loncaric C, Stoos-Veic T, Pejsa V, Jaksic O, Prka Z, Kusec R

(2016) Canonical Wnt/beta-Catenin Signaling Pathway Is Dysregulated in Patients With Primary and

Secondary Myelofibrosis. Clinical lymphoma, myeloma & leukemia 16 (9):523-526.

doi:10.1016/j.clml.2016.06.004

3. Geduk A, Atesoglu EB, Tarkun P, Mehtap O, Hacihanefioglu A, Demirsoy ET, Baydemir C (2015)

Role of beta-Catenin in Bcr/Abl Negative Myeloproliferative Neoplasms: An

Immunohistochemical Study. Clinical lymphoma, myeloma & leukemia 15 (12):785-789.

doi:10.1016/j.clml.2015.08.084

4. Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM, Bloomfield CD, Cazzola

M, Vardiman JW (2016) The 2016 revision to the World Health Organization classification of

myeloid neoplasms and acute leukemia. Blood 127 (20):2391-2405. doi:10.1182/blood-2016-03-

643544

5. Barosi G, Mesa RA, Thiele J, Cervantes F, Campbell PJ, Verstovsek S, Dupriez B, Levine RL,

Passamonti F, Gotlib J, Reilly JT, Vannucchi AM, Hanson CA, Solberg LA, Orazi A, Tefferi A,

International Working Group for Myelofibrosis R, Treatment (2008) Proposed criteria for the

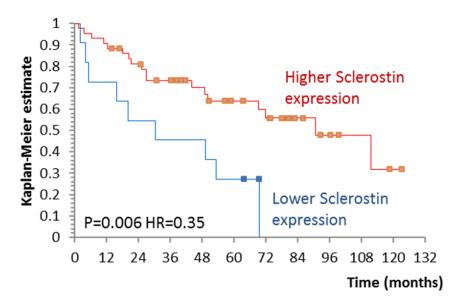
4

diagnosis of post-polycythemia vera and post-essential thrombocythemia myelofibrosis: a consensus statement from the International Working Group for Myelofibrosis Research and Treatment. Leukemia 22 (2):437-438. doi:10.1038/sj.leu.2404914

- 6. Lucijanic M (2016) Survival analysis in clinical practice: analyze your own data using an Excel workbook. Croatian medical journal 57 (1):77-79
- 7. Cain CJ, Rueda R, McLelland B, Collette NM, Loots GG, Manilay JO (2012) Absence of sclerostin adversely affects B-cell survival. Journal of bone and mineral research: the official journal of the American Society for Bone and Mineral Research 27 (7):1451-1461. doi:10.1002/jbmr.1608
- 8. Cervantes F, Dupriez B, Pereira A, Passamonti F, Reilly JT, Morra E, Vannucchi AM, Mesa RA, Demory JL, Barosi G, Rumi E, Tefferi A (2009) New prognostic scoring system for primary myelofibrosis based on a study of the International Working Group for Myelofibrosis Research and Treatment. Blood 113 (13):2895-2901. doi:10.1182/blood-2008-07-170449

Figure: A) Myelofibrosis patients presenting with higher Sclerostin (measured by immunohistochemistry) and B) *SOST* expression (measured by real-time polymerase chain reaction) experienced improved survival. The log-rank test was used.

A) Overall survival by Sclerostin



B) Overall survival by SOST

