Biomarker changes in suspected idiopathic normalpressure hydrocephalus patients undergoing external lumbar drainage: a pilot study

Brgić Mandić, Klara; Mrak, Goran; Barić, Hrvoje; Marasanov, Sergej; Šimić, Goran; Španić Popovački, Ena; Klarica, Marijan

Source / Izvornik: Croatian Medical Journal, 2024, 65, 328 - 338

Journal article, Published version Rad u časopisu, Objavljena verzija rada (izdavačev PDF)

https://doi.org/10.3325/cmj.2024.65.328

Permanent link / Trajna poveznica: https://urn.nsk.hr/um:nbn:hr:105:972063

Rights / Prava: <u>Attribution-NonCommercial-NoDerivatives 4.0 International/Imenovanje-</u> Nekomercijalno-Bez prerada 4.0 međunarodna

Download date / Datum preuzimanja: 2025-03-20



Repository / Repozitorij:

Dr Med - University of Zagreb School of Medicine Digital Repository





328

Biomarker changes in suspected idiopathic normalpressure hydrocephalus patients undergoing external lumbar drainage: a pilot study

Aim To examine whether changes in biomarker concentrations in patients with idiopathic normal-pressure hydrocephalus (iNPH) during 72 h of external lumbar drainage (ELD) can differentiate between responders and non-responders.

Methods Twenty patients with clinical and neuroradiological signs of iNPH underwent ELD over a period of 72 h. During this period, changes in cerebrospinal fluid (CSF) concentrations of biomarkers (amyloid- β , total and phosphorylated tau proteins) and intracranial pressure were monitored, and the volume of drained CSF was measured. Changes in the concentrations of selected biomarkers at three time points (0, 36, and 72 h) during ELD were tested for association with changes in clinical condition.

Results Ten patients showed significant clinical improvement after ELD, quantified as a difference of two or more points on the Mini-Mental State Examination and/or Japanese iNPH grading scale. The concentration of all tested biomarkers increased during the first 36 h. Respondents had higher A β 1-42 at all time points, with a significant difference seen after 72 h. They also had a significantly higher A β 1-42/A β 1-40 ratio at all time points.

Conclusion A gradual increase in $A\beta$ 1-42 concentration during three-day ELD represents a possible positive prognostic factor for the placement of permanent CSF drainage in patients with iNPH.

Klara Brgić Mandić¹, Goran Mrak¹, Hrvoje Barić¹, Sergej Marasanov¹, Goran Šimić², Ena Španić Popovački², Marijan Klarica³

¹Department of Neurosurgery, University Hospital Center Zagreb, Zagreb, Croatia

²Department of Neuroscience, Croatian Institute for Brain Research, University of Zagreb School of Medicine, Zagreb, Croatia

³Department of Pharmacology and Croatian Institute for Brain Research, University of Zagreb, School of Medicine, Zagreb, Croatia

Received: March 1, 2024

Accepted: May 23, 2024

Correspondence to:

Marijan Klarica Department of Pharmacology and Croatian Institute for Brain Research University of Zagreb, School of Medicine Šalata 2 10000 Zagreb, Croatia *mklarica@mef.hr* Idiopathic normal-pressure hydrocephalus (iNPH) is characterized by a triad of symptoms: dementia, gait disturbances, and urinary incontinence (1). Gait disturbances are often the first symptom, present in 94%-100% of patients at diagnosis (2-9). Cognitive impairments are observed in 78%-98% of patients, while urinary urgency and incontinence affect 60%-92% at diagnosis (4,8). The disorder is believed to result from the pressure of enlarged ventricles on motor fibers in the corticospinal pathway (10). The prevalence of iNPH in people older than 65 years is 3.7% (11).

The most common differential diagnosis for iNPH is Alzheimer disease (AD), the most prevalent neurodegenerative condition (12-14). Up to 75% of patients exhibit pathohistological characteristics of both AD and iNPH (14-18). For AD diagnosis, three core biomarkers are crucial: amyloid beta 1-42 (AB 1-42), total tau (t-tau), and phosphorylated tau protein (p-tau) (19,20). The full-length Aβ 1-42 is extremely hydrophobic and forms oligomers and fibrils that accumulate in extracellular plaques, which are characteristic of AD (17). Presumably due to the accumulation of AB 1-42 in plaques, its concentration in cerebrospinal fluid (CSF) is notably lower in AD patients than in the healthy population (21-26). Although in patients with iNPH, amyloid-β concentration tends to be lower than in the healthy population, these values are still higher than those in patients with AD (27). Patients with iNPH have a higher concentration of total tau protein (28), p-tau protein, and amyloid precursor protein (APP) and its fragments in CSF than the healthy population, but lower than AD patients (29). Another useful marker in discriminating iNPH from AD may be CSF phosphorylated tau protein at threonine position 181 (pT181), alone or in combination with total tau (30,31).

Factors determining CSF movement along the CSF system, which affect its interaction with interstitial fluid, certainly influence the fate of molecules of different molecular weight and their distribution between tissue and CSF (32-34). Healthy people, for instance, have a higher concentration of monoamines and their metabolites in cranial than in lumbar CSF, a finding that challenges the traditional view of a unidirectional CSF circulation (35,36). This suggests a more complex interaction between CSF and brain tissue metabolism. When 10 mL (37) to 15 mL (38) of lumbar CSF was sampled, the concentration of various monoamines exponentially increased in individual fractions of the CSF sample (first, middle, and last milliliters). This observation indicates that CSF closer to the brain tissue contains a higher concentration of monoamines and their metabolites, contrary to an even distribution expected according to the classic concept of one-way CSF circulation, reflecting changes in the metabolism of adjacent tissue (37). In addition, in CSF samples obtained by free cisternal drainage in animals over two hours, the concentration of monoamine metabolites exponentially increased, which indicated an influx of CSF from higher parts containing higher concentrations of the measured substances (39).

Contrary to the concentrations of monoamines within the CSF system, the concentrations of blood-derived proteins in the ventricle are lower compared with the lumbar compartment (40,41). When extracting CSF through lumbar puncture in healthy people, the protein concentration in CSF decreases in subsequent fractions (37). This observation additionally indicates that CSF is not mixed by circulation. Moreover, extracting a CSF sample from higher parts with a lower protein concentration can easily explain the observed phenomenon of protein concentration drop in lumbar CSF in later fractions.

iNPH is characterized by biochemical changes in CSF that reflect metabolic changes in the brain. CSF is in direct contact with the extracellular space and is therefore considered a good source of potential biomarkers. Similar to the concentration gradient for proteins in the CSF system, there also appears to be a concentration gradient for peptides such as A β 1-42 (42). The preoperative concentration of A β 1-42 in the lumbar region was shown to be higher than the postoperative concentration in the ventricles (42).

In contrast to healthy people, patients with AD and iNPH are expected to have lower AB 1-42 levels in initial CSF samples due to amyloid accumulation in the interstitial space. Specifically, during preoperative testing, patients with suspected iNPH undergoing prolonged external lumbar drainage (ELD) with larger CSF volumes drained, are expected to have an increased peptide biomarker concentration. This is because low CSF pressure may induce hydrostatic drawing of water from the blood into the interstitial space and CSF, resulting in the "washing out" of accumulated Aβ 1-42 (43,44). We hypothesized that changes in peptide concentration in the CSF of patients evaluated for potential iNPH management differed from those observed previously in healthy people. Furthermore, we postulated that Aβ concentration from the onset to the end of prolonged CSF drainage, due to suspected iNPH, varied between responders and non-responders. Consequently, we assessed AD biomarker concentrations in collected CSF samples during extended ELD immediately after placement, and after 36 and 72 hours, while monitoring changes in CSF pressure, volume of drained CSF, and clinical response.

PATIENTS AND METHODS

Patients

This study was conducted at the University Hospital Center Zagreb from February 2018 to April 2023. The research was approved by the Ethics Committee of the University Hospital Center Zagreb and the Ethics Committee of the University of Zagreb School of Medicine.

The study enrolled patients who underwent drainage placement due to suspected iNPH manifesting as cogni-

tive impairment, gait disturbances, and urinary incontinence. To quantify cognitive impairment, the Mini-Mental Status Examination (MMSE) was used. The Japanese iNPH scale (JiNPHS) was used to quantify other symptoms. A MMSE score of less than 24 indicated significant cognitive deterioration (45).

Patients were required to meet the neuroradiological criteria of iNPH, including ventriculomegaly, especially of the frontal and temporal horns, Evans index >0.3, bulging of the corpus callosum more cranially, callosal angle <90°, significant expansion of the Sylvian fissure disproportionate to the expansion of the convexity sulcus (especially in the parietal region), and increased signal in T2-measured periventricular MRI sequences. In patients with clinical and radiological sus-



FIGURE 1. Flow diagram of patient selection. MRI – magnetic resonance imaging; iNPH – idiopathic normal-pressure hydrocephalus; MMSE – Mini-mental State Examination; JiNPHS – Japanese iNPH Scale; CSF – cerebrospinal fluid.

331

picion of iNPH, standard testing involved placing an external lumbar drain and draining a large amount of CSF (~10 mL/h) over a 72-hour period. The exclusion criteria were underlying neurological or non-neurological disorders that can cause the same symptoms and a history of severe brain trauma. The opening CSF pressure during ELD placement had to be within reference values, meaning $\leq 20 \text{ cm H}_2$ O. Before drainage placement, patients were thoroughly evaluated for the severity of clinical symptoms (MMSE and Japanese iNPH scale). A follow-up clinical evaluation was performed after 72 hours (Figure 1). Patients who had shunt surgery were followed up at 1-month and 6-month intervals postoperatively. Nine patients remained clinically stable, while one required shunt revision due to infection.

Method of CSF pressure recording during ELD

Lumbar drainage was placed between the intervertebral space L_N/L_v or L_VS_μ depending on the anatomical predisposition of the patients, and was determined in relation to the iliac crest. The puncture site was prepared in the standard way, and a 14-16 G Tuohy needle was inserted. After removing the stylet, the needle was connected to the monitor via a transducer placed at the level of the internal auditory canal (IAC), and the first lumbar opening pressure was measured. Subsequently, a lumbar drain (Medtronic EDM Lumbar Drainage Kit, Minneapolis, MN, USA) was placed through the needle and connected to the transducer and the CSF collector via the T connection. CSF pressure was monitored hourly in the horizontal position, corresponding to intracranial pressure when the pressure transducer is positioned at the level of the IAC.

Quantification of biomarkers in CSF

Immediately after ELD placement, 1-2 mL of CSF was sampled for biomarker quantification (β -amyloid proteins [A β

1-42, Aβ 1-40], total tau [t-tau], and phosphorylated-tau [ptau]). Sampling was repeated after 36 h and 72 h. CSF samples were stored in Eppendorf pure polypropylene tubes (Eppendorf, Merck KGaA, Darmstadt, Germany). After centrifugation at 2000×g for 10 min, CSF samples were stored at -80 °C until further analysis. The biomarker levels in CSF were determined with enzyme-linked immunosorbent assay according to the manufacturer's protocols: Aβ1-42 with Innotest β-amyloid 1-42; total tau with Innotest hTau Ag; and p-tau181 with Innotest Phospho-Tau (181P) (all by Fujirebio, Gent, Belgium).

Since biomarker concentrations can sometimes vary significantly (46), each sample was analyzed in duplicate. Each biomarker was measured on the same day using the same batch of reagents. If the measured concentrations of a sample differed by more than 10%, the measurement was repeated. All reported concentrations are the mean values of two measurements.

Statistical analysis

Continuous variables are presented as median (range or interquartile range), and nominal variables as absolute (relative) frequency. Differences between the groups in continuous variables were assessed with the Mann-Whitney U test. Differences between the groups in nominal variables were assessed with the χ^2 test. The before- and after-ELD placement comparisons were performed with the Wilcox-on signed-rank test. Multiple comparisons of biomarker levels across time were performed with the Friedman test. All tests were two-sided. The level of statistical significance was set at P=0.05. SPSS, version 25.0 (IBM Corp., Armonk, NY, USA), and GraphPad Prism, version 8.4.3 (GraphPad Software, Boston, MA, USA), were used for statistical analysis and graphical presentation of the results.

TABLE 1. Sociodemographic and clinical data for responder and non-responder groups. Numbers are median (range) or absolute (relative) frequency*

	Responders (n = 10)		Non-respon	ders (n=10)	_
	median	range	median	range	Р
Age (years)	71	65-82	74	67-82	0.393
Sex (female/male)	5/5	50/50	7/3	70/30	0.650
MMSE before ELD	27	21-28	24	4-29	0.280
MMSE after ELD	29	23-30	26	4-30	0.143
JiNPHS before ELD	5	3-7	4	3-11	>0.999
JiNPHS after ELD	2	1-5	4	1-11	0.015
CSF volume drained in 72 h (mL)	730	570-931	787	650-982	0.278

*Abbreviations: MMSE – Mini-Mental State Examination; JiNPHS – Japanese iNPH Scale; ELD – external lumbar drainage, CSF – cerebrospinal fluid.

RESULTS

A total of 20 patients with suspected iNPH underwent testing. Sixty percent were female, with a median age of 72.5 years (Table 1). This aligns with global findings that iNPH predominantly affects individuals over 65 years of age. MMSE scores significantly increased, and JiNPHS scores significantly decreased after ELD testing (the Wilcoxon signed-rank test) (Table 2).

Out of all patients tested for suspected iNPH, 10 showed significant clinical improvement after ELD (responders), quantified as a difference of two or more points in either MMSE or JiNPHS. Over three days, the average CSF volume drained was 770 mL, ranging from 570 to 982 mL. In the responder group, an average of 730 mL of CSF was drained over 72 hours, compared with 787 mL in the non-responder group (Table 1).

The average opening pressure during ELD placement was 6.5 cm H_2O in both groups. After 36 h, the median pressure in the responder group increased to 7.5 cm H_2O , while it averaged 5 cm H_2O in the non-responder group. The pressure difference between the groups decreased after 72 h (6 cm H_2O and 5 cm H_2O , respectively).

The levels of A β 1-42, A β 1-40, total tau, and pT181 biomarkers significantly changed over time, with the exception

of the A β 1-42/A β 1-40 ratio (Table 3). A β 1-42 levels were higher in responders compared with non-responders at all measured points, with a significant difference noted only after 72 h (Figure 2A, Table 4). Although non-responders had higher total tau (Figure 2C) and A β 1-40 levels (Figure 2B) at all time points, the differences were not significant. Similarly, pT181 levels were higher in the responder group at all times, but the difference did not reach significance (Figure 2D). Responders had a significantly higher A β 1-42/ A β 1-40 ratio at all time points (Figure 2E, Table 4).

DISCUSSION

In our study, the concentration of all tested biomarkers increased within the first 36 hours after drainage placement. This finding suggests that significant CSF volume drainage induces a continuous influx of these substances from tissues (where they accumulate) into CSF. This phenomenon, previously observed during hourly CSF collection over 36 h (47), remains unexplained.

A β 1-42 concentration consistently rose in the responder group, which indicates that continuous CSF drainage facilitates the washout of accumulated A β 1-42 from the interstitial space into CSF. Given that A β 1-42 deposits predominantly in the gray matter, an increase in lumbar CSF A β 1-42 concentration likely results from CSF arriving from cranial regions with higher A β 1-42 concentrations. After

		Ν	Mean rank	Sum of ranks	P value
Mini-mental State Examination	negative ranks	0	0	0	< 0.001
	positive ranks	16	8.50	136.00	
	ties	4			
	total	20			
Japanese Idiopathic Normal-Pressure	negative ranks	13	7.00	91.00	0.001
Hydrocephalus Scale	positive ranks	0	0	0	
	ties	7			
	total	13	7.00	91.00	

TABLE 2. Comparisons of clinical scores before and after external lumbar drainage placement

TABLE 3. Concentrations of amyloid A β 1-42, A β 1-40, total tau protein phosphorylated tau protein (pT181), and amyloid A β 1-42/ A β 1-40 ratio in cerebrospinal fluid immediately after external lumbar drainage placement (0`), after 36 hours, and after 72 hours of drainage

	0`		36 h		72 h		_
	median	range	median	range	median	range	Р
Aβ1-42 (pg/mL)	144.47	42.86-1486.14	320.02	15.24-1386.78	273.93	42.86-580.41	0.010
Aβ1-40 (pg/mL)	2531.05	341.13-14250.56	4181.17	1064.94-22197.72	4254.3	384.28-21188.43	0.027
Aβ1-42/Aβ1-40 (pg/mL)	0.08424	0.02381-0.22653	0.06166	0.00686-0.22069	0.05726	0.01927-0.23922	<0.911
Total tau	161.2	22.69-334.99	275.8	77.51-693.76	296.33	15.77-1027.10	< 0.001
Tau pT181	27.42	13.14-58.86	50.92	23.57-108.78	48.74	17.83-123.23	< 0.001

72 hours, $A\beta$ 1-42 levels were significantly higher in the responder group, which suggests this could be a new prognostic factor for surgery if confirmed in larger studies.

CSF biomarkers are used to differentiate patients likely to show clinical improvement after the placement of a permanent shunt from those mimicking iNPH symptoms. Leinonen et al demonstrated that in 22% of patients with suspected iNPH, the presence of amyloid plaques and neurofibrillary tangles in biopsy samples correlated with AD development over 4.4 years (18). AD is characterized by an increased concentration of total and phosphorylated tau proteins and a decreased concentration of AB 1-42 (20,21). A meta-analysis of 13 studies (48-60) showed that shunt-responder patients had lower lumbar CSF concentrations of total and phosphorylated tau proteins than non-responders, with no significant difference in A β 1-42 levels (61). However, the generalizability of these results is limited due to variations in analytical methods, methodological weaknesses of the studies, the small number of studies dealing with AB 1-42 changes, and differing CSF sample collection times after varying drainage durations. Tarnaris et al demonstrated, in 11 patients with suspected iNPH, that the concentrations of AB1-42 and total tau protein increased during 72 h of CSF evacuation via ELD (62). Jingami et al found that during the tap test, there was an increased concentration of total tau protein and decreased levels of A β 1-42 in the last milliliter compared with the first milliliter of CSF (63). Considering that the volume of drained CSF in the cited study was 30 mL, these results cannot be directly compared with ours, where the drained CSF volume was considerably higher (770 mL).

Our findings demonstrate changes at three time points (0', 36 h, 72 h) during ELD of a significant total CSF volume (average 770 mL under lower CSF pressure). In the responder group, the concentration of the AB 1-42 isoform was consistently higher at all time points compared with the nonresponder group, with the difference reaching significance only after 72 h. This observation, under these specific conditions, significantly differs from those of previous studies. Additionally, in our study, the non-responder group had higher total tau protein and AB 1-40 at all time points, though without reaching statistical significance. In the responder group, the concentration of total tau protein decreased after 72 h, which indicated that following the initial washout of this marker characteristic of neurofibrillary neurodegeneration, its concentration in patients without AD comorbidity in the lumbar CSF sample stabilizes. Higher values of pT181 were also observed in the responder

TABLE 4. Concentrations of amyloid $A\beta$ 1-42, $A\beta$ 1-40, total tau protein, phosphorylated tau protein (pT181), and amyloid $A\beta$ 1-42 / $A\beta$ 1-40 ratio in cerebrospinal fluid in the responder and non-responder groups immediately after external lumbar drainage placement (0'), after 36 hours, and after 72 hours of drainage

	Responders (n=10)		Non-re		
	median	range	median	range	P
Aβ1-42 (pg/mL)					
0`	224.535	71.99-1486.14	122.285	42.86-481.24	0.579
36 h	373.5	183.8-1386.78	242.435	15.24-409.89	0.052
72 h	476.52	96.19-580.41	168.755	42.86-391.52	0.043
Aβ1-40 (pg/mL)					
0`	1668.76	384.28-14250.56	3621.33	341.13-11462.74	0.436
36 h	3336.72	1137.97-22197.72	5164.27	1064.94-8969.21	0.796
72 h	2756.75	424.6-21188.43	4721.04	384.28-10134.47	0.720
Αβ1-42/Αβ1-40					
0`	0.14183	0.03153-0.22653	0.05037	0.02381-0.12564	0.007
36 h	0.11842	0.02858-0.21062	0.04065	0.00686-0.22069	0.019
72 h	0.16955	0.02529-0.23922	0.04845	0.01927-0.11153	0.008
t- tau (pg/mL)					
0`	150.85	55.72-334.99	163.24	22.69-331.34	0.796
36 h	257.905	138.1-420.49	319.77	77.51-693.76	0.481
72 h	235.9	55.72-455.39	339.1	15.77-1027.1	0.400
pT181 (pg/mL)					
0`	38.05	13.14-58.86	26.77	15.21-44.77	0.481
36 h	47.15	30.90-108.78	44.06	23.57-107.53	0.912
72 h	56.26	17.83-105.27	44.71	22.31-123.23	0.604

group at all time points, but without a significant difference, which contrasts with the findings of previous studies, in which responders had lower values of pT181.

Another longitudinal study, tracking the concentrations of biomarkers in the lumbar and ventricular spaces over time, showed that in patients with probable iNPH and a negative A β brain biopsy, the concentration of A β 1-42 shifted toward more positive values compared with patients with iNPH and a positive A β brain biopsy, particularly after a longer follow-up. This indicates that, in patients with probable iNPH, a higher A β 1-42 value acts as the best negative predictor of underlying AD (64). These findings support the notion of a better clinical response to permanent shunt placement in patients with higher A β 1-42 levels in the lumbar CSF, aligning well with our results.

According to the new concept of CSF physiology, CSF can be produced at the level of the brain and spinal tissue capillary network if an osmotic or hydrostatic gradient is created, which facilitates the entry of a net volume of fluid from the capillaries into the interstitial space and CSF (43,44). During ELD, the reduction of hydrostatic CSF pressure to 5-7.5 cm H_2O enables the entry of fluid from the capillary network into the interstitial space of the cerebral gray matter and potentially facilitates the "washout" of substances accumulated in the interstitium into the CSF system. This mechanism may explain the observed gradual increase in A β 1-42 concentration during prolonged ELD in responders.

The current study is subject to several major limitations, including a small sample size, single-center design, and the absence of long-term follow-up data. These factors limit the generalizability of our findings and the ability to assess how biomarker changes correlate with long-term outcomes, such as the progression of neurological symptoms and mortality. Additionally, the lack of a control group to directly compare the biomarker levels may also affect the interpretation of our results. Without a control group, it is challenging to ascertain whether the observed biomarker



FIGURE 2. Concentrations of different biomarkers in the responder and non-responder groups: (**A**) amyloid A β 1-42, (**B**) A β 1-40, (**C**) total tau protein (t-tau), (**D**) phosphorylated tau protein (pT181) (expressed in pg/mL), and (**E**) A β 1-42 /A β 1-40 ratio in cerebrospinal fluid at time points: 0[°] – immediately after external lumbar drainage placement, after 36 hours, and after 72 hours of drainage; * significant difference.

changes are specific to the treatment received or reflect natural disease progression. Another potential limitation is the variability in the diagnostic criteria for iNPH across different centers, which may introduce selection bias and affect the applicability of our findings to a broader population. Furthermore, we did not perform a brain biopsy, which would definitively determine the coexistence of an underlying neurodegenerative disease. Finally, our study did not account for potential confounding factors such as variations in patients' medication use, comorbid conditions, or lifestyle factors that could influence biomarker levels. Addressing these limitations in future research is essential for a more comprehensive understanding of the implications of biomarker changes in patients with iNPH.

Funding This work was supported by the Croatian Science Foundation and the Ministry of Science and Education of the Republic of Croatia (Projects: 1. Pathophysiology of Cerebrospinal Fluid and Intracranial Pressure. No. 108-1080231-0023; 2. Cerebrospinal Fluid Volume and Pressure Regulation. No. 10106-20-2502; 3. Role of Blood-Brain Barrier, Innate Immunity, and Tau Protein Oligomerization in the Pathogenesis of Alzheimer's Disease." No. CSF IP-2019-04-3584). Research was co-financed by the Scientific Centre of Excellence for Basic, Clinical, and Translational Neuroscience CORE-NEURO (project Experimental and Clinical Research of Hypoxic–Ischemic Damage in Perinatal and Adult Brain; GA KK01.1.101.0007 funded by the European Union through the European Regional Development Fund).

Ethical approval granted by the the Ethics Committee of the Clinical Hospital Center Zagreb (Class: 8.1-17/202-2, number: 02/21 AG), and by the Ethics Committee of the University of Zagreb School of Medicine (Class: 641-01/23-02/01, number: 251-59-10106-23-111/213).

Declaration of authorship KBM, GM, MK conceived and designed the study; KBM, HB, SMM acquired the data; KBM, GM, HB, GŠ, EŠP, MK analyzed and interpreted the data; KBM, MK drafted the manuscript; all authors critically reviewed the manuscript for important intellectual content; all authors gave approval of the version to be submitted; all authors agree to be accountable for all aspects of the work.

Competing interests HB is a Deputy Editor-in-Chief of the *Croatian Medical Journal*. To ensure that any possible conflict of interest relevant to the journal has been addressed, this article was reviewed according to best practice guidelines of international editorial organizations. All authors have completed the Unified Competing Interest form at www.icmje.org/coi_disclosure.pdf (available on request from the corresponding author) and declare: no support from any organizations that might have an interest in the submitted work in the previous 3 years; no other relationships or activities that could appear to have influenced the submitted work.

References

- 1 Hakim CA, Hakim R, Hakim S. Normal-pressure hydrocephalus. Neurosurg Clin N Am. 2001;12:761-73. Medline:11524297 doi:10.1016/S1042-3680(18)30033-0
- 2 Black PM. Idiopathic normal-pressure hydrocephalus. Results of shunting in 62 patients. J Neurosurg. 1980;52:371-7. Medline:7359191 doi:10.3171/jns.1980.52.3.0371
- 3 Marmarou A, Bergsneider M, Relkin N, Klinge P, Black PM. Development of guidelines for idiopathic normal-pressure hydrocephalus: introduction. Neurosurgery. 2005;57:S1-3. Medline:16160424 doi:10.1227/01.NEU.0000168188.25559.0E

- 4 Miyoshi N, Kazui H, Ogino A, Ishikawa M, Miyake H, Tokunaga H, et al. Association between cognitive impairment and gait disturbance in patients with idiopathic normal pressure hydrocephalus. Dement Geriatr Cogn Disord. 2005;20:71-6. Medline:15908748 doi:10.1159/000085858
- 5 Stolze H, Kuhtz-Buschbeck JP, Drücke H, Jöhnk K, Diercks C, Palmié S, et al. Gait analysis in idiopathic normal pressure hydrocephalus which parameters respond to the CSF tap test? Clin Neurophysiol. 2000;111:1678-86. Medline:10964082 doi:10.1016/S1388-2457(00)00362-X
- 6 Stolze H, Kuhtz-Buschbeck JP, Drücke H, Jöhnk K, Illert M, Deuschl G. Comparative analysis of the gait disorder of normal pressure hydrocephalus and Parkinson's disease. J Neurol Neurosurg Psychiatry. 2001;70:289-97. Medline:11181848 doi:10.1136/ jnnp.70.3.289
- 7 Williams MA, Thomas G, de Lateur B, Imteyaz H, Rose JG, Shore WS, et al. Objective assessment of gait in normal-pressure hydrocephalus. Am J Phys Med Rehabil. 2008;87:39-45. Medline:17993991 doi:10.1097/PHM.0b013e31815b6461
- Iddon JL, Morgan DJ, Loveday C, Sahakian BJ, Pickard JD.
 Neuropsychological profile of young adults with spina bifida with or without hydrocephalus. J Neurol Neurosurg Psychiatry.
 2004;75:1112-8. Medline:15258211 doi:10.1136/jnnp.2003.029058
- 9 Boon AJ, Tans JT, Delwel EJ, Egeler-Peerdeman SM, Hanlo PW, Wurzer HA, et al. Dutch normal-pressure hydrocephalus study: prediction of outcome after shunting by resistance to outflow of cerebrospinal fluid. J Neurosurg. 1997;87:687-93. Medline:9347976 doi:10.3171/jns.1997.87.5.0687
- 10 Hattingen E, Jurcoane A, Melber J, Blasel S, Zanella FE, Neumann-Haefelin T, et al. Diffusion tensor imaging in patients with adult chronic idiopathic hydrocephalus. Neurosurgery. 2010;66:917-24. Medline:20404696 doi:10.1227/01.NEU.0000367801.35654.EC
- 11 Andersson J, Rosell M, Kockum K, Lilja-Lund O, Söderström L, Laurell K. Prevalence of idiopathic normal pressure hydrocephalus: a prospective, population-based study. PLoS One. 2019;14:e0217705. Medline:31141553 doi:10.1371/journal. pone.0217705
- 12 Skalicky P, Mladek A, Vlasak A, De Lacy P, Benes V, Bradac O. Normal pressure hydrocephalus-an overview of pathophysiological mechanisms and diagnostic procedures. Neurosurg Rev. 2020;43:1451-64. Medline:31705404 doi:10.1007/s10143-019-01201-5
- Siraj S. An overview of normal pressure hydrocephalus and its importance: How much do we really know? J Am Med Dir Assoc.
 2011;12:19-21. Medline:21194654 doi:10.1016/j.jamda.2010.05.005
- 14 Golomb J, Wisoff J, Miller DC, Boksay I, Kluger A, Weiner H, et al. Alzheimer's disease comorbidity in normal pressure hydrocephalus: Prevalence and shunt response. J Neurol Neurosurg Psychiatry. 2000;68:778-81. Medline:10811706 doi:10.1136/jnnp.68.6.778

335

- 15 Del Bigio MR, Cardoso ER, Halliday WC. Neuropathological changes in chronic adult hydrocephalus: Cortical biopsies and autopsy findings. Can J Neurol Sci. 1997;24:121-6. Medline:9164688 doi:10.1017/S0317167100021442
- Savolainen S, Paljarvi L, Vapalahti M. Prevalence of Alzheimer's disease in patients investigated for presumed normal pressure hydrocephalus: A clinical and neuropathological study.
 Acta Neurochir (Wien). 1999;141:849-53. Medline:10536721 doi:10.1007/s007010050386
- 17 Bech-Azeddine R, Hogh P, Juhler M, Gjerris F, Waldemar G. Idiopathic normal-pressure hydrocephalus: Clinical comorbidity correlated with cerebral biopsy findings and outcome of cerebrospinal fluid shunting. J Neurol Neurosurg Psychiatry. 2007;78:157-61. Medline:17012342 doi:10.1136/jnnp.2006.095117
- 18 Leinonen V, Alafuzoff I, Aalto S, Suotunen T, Savolainen S, Nagren K, et al. Assessment of beta-amyloid in a frontal cortical brain biopsy specimen and by positron emission tomography with carbon 11labeled Pittsburgh Compound B. Arch Neurol. 2008;65:1304-9. Medline:18695050 doi:10.1001/archneur.65.10.noc80013
- 19 Babić M, Švob Štrac D, Mück-Šeler D, Pivac N, Stanić G, Hof PR, et al. Update on the core and developing cerebrospinal fluid biomarkers for Alzheimer's disease. Croat Med J. 2014;55:347-65. Medline:25165049 doi:10.3325/cmj.2014.55.347
- 20 Frisoni GB, Festari C, Massa F, Ramusino MC, Orini S, Aarsland D, et al. European intersocietal recommendations for the biomarker-based diagnosis of neurocognitive disorders. Lancet Neurol. 2024;23:302-12. Medline:38365381 doi:10.1016/S1474-4422(23)00447-7
- 21 Formichi P, Battisti C, Radi E, Federico A. Cerebrospinal fluid tau, Aβ, and phosphorylated tau protein for the diagnosis of Alzheimer's disease. J Cell Physiol. 2006;208:39-46. Medline:16447254 doi:10.1002/jcp.20602
- 22 Lazo ND, Maji SK, Fradinger EA, Bitan G, Teplow DB. The amyloid β-protein. U: Sipe JD. Amyloid proteins - the beta sheet conformation and disease. Weinheim: Wiley-VCH; 2005: p. 385-491.
- 23 Fagan AM, Roe CM, Xiong C, Mintun MA, Morris JC, Holtzman DM. Cerebrospinal fluid tau/β-amyloid 42 ratio as a prediction of cognitive decline in nondemented older adults. Arch Neurol. 2007;64:343-9. Medline:17210801 doi:10.1001/ archneur.64.3.noc60123
- 24 Andreasen N, Hesse C, Davidsson P, Minthon L, Wallin A, Winblad B, et al. Cerebrospinal fluid β-amyloid(1-42) in Alzheimer disease: differences between early- and late-onset Alzheimer disease and stability during the course of disease. Arch Neurol. 1999;56:673-80. Medline:10369305 doi:10.1001/archneur.56.6.673
- 25 Galasko D, Chang L, Motter R, Clark CM, Kaye J, Knopman D, et al. High cerebrospinal fluid tau and low amyloid β42 levels in the clinical diagnosis of Alzheimer disease and relation to apolipoprotein E genotype. Arch Neurol. 1998;55:937-45. Medline:9678311 doi:10.1001/archneur.55.7.937

- Hampel H, Teipel SJ, Fuchsberger T, Andreasen N, Wiltfang J, Otto M, et al. Value of CSF β-amyloid1-42 and tau as predictors of Alzheimer's disease in patients with mild cognitive impairment. Mol Psychiatry. 2004;9:705-10. Medline:14699432 doi:10.1038/ sj.mp.4001473
- 27 Thal LJ, Kantarci K, Reiman EM, Klunk WE, Weiner MW, Zetterberg H, et al. The role of biomarkers in clinical trials for Alzheimer disease. Alzheimer Dis Assoc Disord. 2006;20:6-15. Medline:16493230 doi:10.1097/01.wad.0000191420.61260.a8
- 28 Kudo T, Mima T, Hashimoto R, Nakao K, Morihara T, Tanimukai H, et al. Tau protein is a potential biological marker for normal pressure hydrocephalus. Psychiatry Clin Neurosci. 2000;54:199-202. Medline:10803815 doi:10.1046/j.1440-1819.2000.00658.x
- 29 Miyajima M, Nakajima M, Ogino I, Miyata H, Motoi Y, Arai H. Soluble amyloid precursor protein alpha in the cerebrospinal fluid as a diagnostic and prognostic biomarker for idiopathic normal pressure hydrocephalus. Eur J Neurol. 2013;20:236-42. Medline:22672777 doi:10.1111/j.1468-1331.2012.03781.x
- 30 Kapaki EN, Paraskevas GP, Tzerakis NG, Sfagos C, Seretis A, Kararizou E, et al. Cerebrospinal fluid tau, phosho-tau181 and beta-amyloid1-42 in idiopathic normal-pressure hydrocephalus: a discrimination from Alzheimer's disease. Eur J Neurol. 2007;14:168-73. Medline:17250725 doi:10.1111/j.1468-1331.2006.01593.x
- 31 Graff-Radford NR. Alzheimer CSF biomarkers may be misleading in normal-pressure hydrocephalus. Neurology. 2014;83:1573-5. Medline:25332445 doi:10.1212/WNL.00000000000916
- 32 Klarica M, Radoš M, Orešković D. The movement of cerebrospinal fluid and its relationship with substances behavior in cerebrospinal and interstitial fluid. Neuroscience. 2019;414:28-48. Medline:31279048 doi:10.1016/j.neuroscience.2019.06.032
- 33 Mestre H, Verma N, Greene TD, Lin LA, Ladron-de-Guevara A, Sweeney AM, et al. Periarteriolar spaces modulate cerebrospinal fluid transport into brain and demonstrate altered morphology in aging and Alzheimer's disease. Nat Commun. 2022;13:3897. Medline:35794106 doi:10.1038/s41467-022-31257-9
- Coto-Vilcapoma MA, Castilla-Silgado J, Fernández-García B,
 Pinto-Hernández P, Cipriani R, Capetillo-Zarate E, et al. New, fully
 implantable device for selective clearance of CSF-target molecules:
 proof of concept in a murine model of alzheimer's disease.
 Int J Mol Sci. 2022;23:9256. Medline:36012525 doi:10.3390/
 ijms23169256
- 35 Bulat M, Živković B. Origin of 5-hydroxyindoleacetic acid in the spinal fluid. Science. 1971;173:738-40. Medline:5109594 doi:10.1126/science.173.3998.738
- 36 Malm J, Kristensen B, Ekstedt J, Wester P. CSF concentration gradients of monoamine metabolites in patients with hydrocephalus. Neurol Neurosurg Psychiatry. 1994;57:1026-33. Medline:7522267 doi:10.1136/jnnp.57.9.1026
- 37 Tigchelaar C, Muller WD, Atmosoerodjo SD, Wardenaar KJ, Kema IP, Absalom AR, et al. Concentration gradients of monoamines, their

precursors and metabolites in serial lumbar cerebrospinal fluid of neurologically healthy patients determined with a novel LC-MS/ MS technique. Fluids Barriers CNS. 2023;20:13. Medline:36782208 doi:10.1186/s12987-023-00413-8

- 38 Jakupčević M, Lacković Z, Stefoski D, Bulat M. Nonhomogeneous distribution of 5-hydroxyindoleacetic acid and homovanillic acid in the lumbar cerebrospinal fluid of man. J Neurol Sci. 1977;31:165-71. Medline:839230 doi:10.1016/0022-510X(77)90103-4
- 39 Maraković J, Vukić M, Radoš M, Chudy D, Klarica M, Orešković D. Monoamine neurotransmitter metabolite concentration as a marker of cerebrospinal fluid volume changes. Acta Neurochir Suppl (Wien). 2016;122:283-6. Medline:27165922 doi:10.1007/978-3-319-22533-3_56
- 40 Fishman RA, Ransohoff J, Osserman EF. Factors influencing the concentration gradient of protein in cerebrospinal fluid. Annual Meeting of the American Neurological Association. 1955;1419-24.
- 41 Wilhelmy F, Krause M, Schob S, Merkenschlager A, Wachowiak R, Härtig W, et al. Cerebrospinal fluid protein concentrations in hydrocephalus. Children (Basel). 2023;10:644. Medline:37189895
- 42 Lukkarinen H, Vanninen A, Tesseur I, Pemberton D, Van Der Ark P, Kokkola T, et al. Concordance of Alzheimer's disease-related biomarkers between intraventricular and lumbar cerebrospinal fluid in idiopathic normal pressure hydrocephalus. J Alzheimers Dis. 2023;91:305-19. Medline:36404546 doi:10.3233/JAD-220652
- 43 Bulat M, Klarica M. Recent insights into a new hydrodynamics of the cerebrospinal fluid. Brain Res Brain Res Rev. 2011;65:99-112. Medline:20817024 doi:10.1016/j.brainresrev.2010.08.002
- Orešković D, Klarica M. The formation of cerebrospinal fluid: nearly a hundred years of interpretations and misinterpretations. Brain Res Brain Res Rev. 2010;64:241-62. Medline:20435061 doi:10.1016/j.brainresrev.2010.04.006
- 45 Boban M, Malojčić B, Mimica N, Vuković S, Zrilić I, Hof PR, et al. The reliability and validity of the Mini-Mental State Examination in the elderly Croatian population. Dement Geriatr Cogn Disord. 2012;33:385-92. Medline:22814030 doi:10.1159/000339596
- 46 Babić M, Vogrinc Ž, Diana A, Klepac N, Borovečki F, Hof PR, et al. Comparison of two commercial enzyme-linked immunosorbent assays for cerebrospinal fluid measurement of amyloid β1-42 and total tau. Transl Neurosci. 2013;4:234-40. Medline:24376914 doi:10.2478/s13380-013-0123-4
- Slats D, Claassen J, Spies PE, Borm G, Besse KTC, van Aalst W, et al. Hourly variability of cerebrospinal fluid biomarkers in Alzheimer's disease subjects and healthy older volunteers. Neurobiol Aging. 2012;33:831-9. Medline:21880396 doi:10.1016/j. neurobiolaging.2011.07.008
- 48 Agren-Wilsson A, Lekman A, Sjoberg W, Rosengren L, Blennow K, Bergenheim AT, et al. CSF biomarkers in the evaluation of idiopathic normal pressure hydrocephalus. Acta Neurol Scand. 2007;116:333-9. Medline:17922727 doi:10.1111/j.1600-0404.2007.00890.x

- 49 Tullberg M, Blennow K, Mansson JE, Fredman P, Tisell M, Wikkelso C. Cerebrospinal fluid markers before and after shunting in patients with secondary and idiopathic normal pressure hydrocephalus. Cereb Fluid Res. 2008;5:9. Medline:18439296 doi:10.1186/1743-8454-5-9
- 50 Abu Hamdeh S, Virhammar J, Sehlin D, Alafuzoff I, Cesarini KG, Marklund N. Brain tissue Aβ42 levels are linked to shunt response in idiopathic normal pressure hydrocephalus. J Neurosurg. 2018;130:121-9. Medline:29350601 doi:10.3171/2017.7.JNS171005
- 51 Craven CL, Baudracco I, Zetterberg H, Lunn MPT, Chapman MD, Lakdawala N, et al. The predictive value of T-tau and AB1-42 levels in idiopathic normal pressure hydrocephalus. Acta Neurochir (Wien). 2017;159:2293-300. Medline:28889317 doi:10.1007/ s00701-017-3314-x
- 52 Hong YJ, Kim MJ, Jeong E, Kim JE, Hwang J, Lee JI, et al. Preoperative biomarkers in patients with idiopathic normal pressure hydrocephalus showing a favorable shunt surgery outcome. J Neurol Sci. 2018;387:21-6. Medline:29571865 doi:10.1016/i.jns.2018.01.017
- 53 Johansson BB, Wikkelsø C, Blomstrand C, Linder L, Fahrenkrug J. Vasoactive intestinal polypeptide in cerebrospinal fluid of patients with dementia. Nord Psykiatr Tidsskr. 1985;39:69-73. doi:10.3109/08039488509101960
- 54 Migliorati K, Panciani PP, Pertichetti M, Borroni B, Archetti S, Rozzini L, et al. P-Tau as prognostic marker in long term follow up for patients with shunted iNPH. Neurol Res. 2021;43:78-85. Medline:33059546 doi:10.1080/01616412.2020.1831300
- 55 Minta K, Jeppsson A, Brinkmalm G, Portelius E, Zetterberg H, Blennow K, et al. Lumbar and ventricular CSF concentrations of extracellular matrix proteins before and after shunt surgery in idiopathic normal pressure hydrocephalus. Fluids Barriers CNS. 2021;18:23. Medline:33985551 doi:10.1186/s12987-021-00256-1
- 56 Patel S, Lee EB, Xie SX, Law A, Jackson EM, Arnold SE, et al. Phosphorylated tau/amyloid beta 1-42 ratio in ventricular cerebrospinal fluid reflects outcome in idiopathic normal pressure hydrocephalus. Fluids Barriers CNS. 2012;9:7. Medline:22444461 doi:10.1186/2045-8118-9-7
- 57 Pyykkö OT, Helisalmi S, Koivisto AM, Mölsä JA, Rummukainen J, Nerg O, et al. APOE4 predicts amyloid-β in cortical brain biopsy but not idiopathic normal pressure hydrocephalus. J Neurol Neurosurg Psychiatry. 2012;83:1119-24. Medline:22955176 doi:10.1136/jnnp-2011-303849
- 58 Scollato A, Terreni A, Caldini A, Salvadori B, Gallina P, Francese S, et al. CSF proteomic analysis in patients with normal pressure hydrocephalus selected for the shunt: CSF biomarkers of response to surgical treatment. Neurol Sci. 2010;31:283-91. Medline:19936883 doi:10.1007/s10072-009-0181-0
- 59 Tarnaris A, Toma AK, Chapman MD, Keir G, Kitchen ND, Watkins LD. Use of cerebrospinal fluid amyloid-β and total tau protein to predict favorable surgical outcomes in patients with idiopathic

337

normal pressure hydrocephalus. J Neurosurg. 2011;115:145-50. Medline:21438653 doi:10.3171/2011.2.JNS101316

- 60 Vanninen A, Nakajima M, Miyajima M, Rauramaa T, Kokki M, Musialowicz T, et al. Elevated CSF LRG and decreased Alzheimer's disease biomarkers in idiopathic normal pressure hydrocephalus. J Clin Med. 2021;10:1105. Medline:33800840 doi:10.3390/ jcm10051105
- 61 Thavarajasingam SG, El-Khatib M, Vemulapalli KV, Iradukunda HAS, Laleye J, Russo S, et al. Cerebrospinal fluid and venous biomarkers of shunt-responsive idiopathic normal pressure hydrocephalus: a systematic review and meta-analysis. Acta Neurochir (Wien). 2022;164:1719-46. Medline:35230552 doi:10.1007/s00701-022-05154-5
- 62 Tarnaris A, Toma AK, Chapman MD, Petzold A, Kitchen ND, Keir G, et al. The longitudinal profile of CSF markers during external lumbar drainage. J Neurol Neurosurg Psychiatry. 2009;80:1130-3. Medline:19535354 doi:10.1136/jnnp.2008.171686

- 63 Jingami N, Uemura K, Asada-Utsugi M, Kuzuya A, Yamada S, Ishikawa M, et al. Two-Point Dynamic Observation of Alzheimer's Disease Cerebrospinal Fluid Biomarkers in Idiopathic Normal Pressure Hydrocephalus. J Alzheimers Dis. 2019;72:271-7. Medline:31561378 doi:10.3233/JAD-190775
- Lukkarinen H, Tesseur I, Pemberton D, Van Der Ark P, Timmers M,
 Slemmon R, et al. Time Trends of Cerebrospinal Fluid Biomarkers of
 Neurodegeneration in Idiopathic Normal Pressure Hydrocephalus.
 J Alzheimers Dis. 2021;80:1629-42. Medline:33720890 doi:10.3233/
 JAD-201361