

Association of the MAOB rs1799836 Single Nucleotide Polymorphism and APOE ε4 Allele in Alzheimer's Disease

Babić Leko, Mirjana; Nikolac Perković, Matea; Nedić Erjavec, Gordana; Klepac, Nataša; Švob Štrac, Dubravka; Borovečki, Fran; Pivac, Nela; Hof, Patrick R.; Šimić, Goran

Source / Izvornik: **Current Alzheimer Research, 2021, 18, 585 - 594**

Journal article, Published version

Rad u časopisu, Objavljena verzija rada (izdavačev PDF)

<https://doi.org/10.2174/1567205018666210917162843>

Permanent link / Trajna poveznica: <https://um.nsk.hr/um:nbn:hr:105:989394>

Rights / Prava: [In copyright](#)/[Zaštićeno autorskim pravom.](#)

Download date / Datum preuzimanja: **2024-07-13**



Repository / Repozitorij:

[Dr Med - University of Zagreb School of Medicine Digital Repository](#)



ORIGINAL ARTICLE

Association of the *MAOB* rs1799836 Single Nucleotide Polymorphism and *APOE* ϵ 4 Allele in Alzheimer's Disease

Mirjana Babić Leko^a, Matea Nikolac Perković^b, Gordana Nedić Erjavec^b, Nataša Klepac^c, Dubravka Švob Štrac^b, Fran Borovečki^c, Nela Pivac^b, Patrick R. Hof^d, Goran Šimić^{*a}

^aDepartment of Neuroscience, Croatian Institute for Brain Research, University of Zagreb Medical School, Zagreb, Croatia

^bDepartment of Molecular Medicine, Institute Ruđer Bošković, Zagreb, Croatia

^cDepartment of Neurology, University Hospital Centre Zagreb, Zagreb, Croatia

^dNash Family Department of Neuroscience, Friedman Brain Institute, and Ronald M. Loeb Center for Alzheimer's Disease, Icahn School of Medicine at Mount Sinai, New York, USA

Abstract: Background: The dopaminergic system is functionally compromised in Alzheimer's disease (AD). The activity of monoamine oxidase B (MAOB), the enzyme involved in the degradation of dopamine, is increased during AD. Also, increased expression of MAOB occurs in the postmortem hippocampus and neocortex of patients with AD. The *MAOB* rs1799836 polymorphism modulates *MAOB* transcription, consequently influencing protein translation and MAOB activity. We recently showed that the cerebrospinal fluid levels of amyloid β_{1-42} are decreased in patients carrying the A allele in *MAOB* rs1799836 polymorphism. **Objective:** The present study compares *MAOB* rs1799836 polymorphism and *APOE*, the only confirmed genetic risk factor for sporadic AD. **Method:** We included 253 participants, 127 of whom had AD, 57 had mild cognitive impairment, 11 were healthy controls, and 58 suffered from other primary causes of dementia. *MAOB* and *APOE* polymorphisms were determined using TaqMan SNP Genotyping Assays. **Results:** We observed that the frequency of *APOE* ϵ 4/ ϵ 4 homozygotes and *APOE* ϵ 4 carriers is significantly increased among patients carrying the AA *MAOB* rs1799836 genotype. **Conclusions:** These results, together with the results of our previous study, indicate that the *MAOB* rs1799836 polymorphism is a potential genetic biomarker of AD.

ARTICLE HISTORY

Received:
Revised:
Accepted:

DOI:

Key words: Alzheimer's disease, MAOB, APOE, polymorphisms, genetic biomarkers, mild cognitive impairment

1. INTRODUCTION

The only confirmed genetic risk factor for sporadic Alzheimer's disease (AD) that comprises over 99% of all AD cases, is the apolipoprotein E gene (*APOE*) ϵ 4 variant [1,2]. The APOE protein is mainly produced by astrocytes and is involved in transport of cholesterol to neurons [3]. Two single nucleotide polymorphisms (SNP) in the *APOE* gene that affect *APOE* transcription and translation result in three common APOE variants (APOE ϵ 2, ϵ 3 and ϵ 4) [4]. While *APOE* ϵ 2 variant is considered to have protective

effect in AD [5], *APOE* ϵ 4 heterozygotes have a 5-time increased risk, and *APOE* ϵ 4 homozygotes a 20-time increased risk of developing AD [6]. Several other genes have been recently associated with the increased risk for development of sporadic AD, such as *ABCA7*, *BINI*, *CD33*, *CD2AP*, *CLU*, *CRI*, *MS4A6A*, *MS4A4E*, *PICALM* [7–12], *PLD3* [13], *TREM2* [14] among others. However, the influence that these genes have on increasing the risk of AD is far less than that of the *APOE* gene.

Monoamine oxidase B (MAOB) is the enzyme bound to the outer mitochondrial membrane responsible for the degradation of dopamine. MAOB activity is increased in AD [15–18], contributing to decreasing dopamine levels [19–21]. Moreover, it was proved that MAOB inhibitors increase dopamine levels in the brain (reviewed in [22]). Some polymorphisms within the *MAOB* gene can also affect MAOB activity. The *MAOB* rs1799836 SNP (A644G) modifies *MAOB* transcription and translation leading to altered activity [23]. In fact, we recently observed that cerebrospinal fluid (CSF) amyloid β_{1-42} ($A\beta_{1-42}$) levels are decreased in patients carrying the A allele in *MAOB* rs1799836 polymorphism [24]. Here, we assess the potential association of this polymorphism with *APOE* genotype variants in AD.

*Address correspondence to Goran Šimić at School of Medicine, University of Zagreb, Šalata 12, 10 000 Zagreb, Croatia; Tel/Fax: +385-1-459-6807, +385-1-459-6942; E-mail: gsimic@hiim.hr

2. MATERIALS AND METHOD

2.1. Subjects

The study included a total of 253 subjects recruited at the University Hospital Center Zagreb, Croatia. The subjects presented with various types of dementia, including AD, mild cognitive impairment (MCI), vascular cognitive impairment/vascular dementia (VaD), frontotemporal dementia (FTD), dementia with Lewy bodies (DLB), mixed dementia (AD+VaD), corticobasal syndrome (CBS), non-specific dementia (ND) and Parkinson's disease (PD). AD was diagnosed in 127 patients using the criteria of the National Institutes on Aging - Alzheimer's Association (NIA-AA) [25]. MCI was diagnosed in 57 patients using the criteria of Petersen *et al.* [26] and Albert *et al.* [27]. FTD was diagnosed in 25 patients using the criteria of Neary *et al.* [28], while VaD was diagnosed in 15 patients using the criteria of National Institute for Neurological Disorders and Stroke - Association Internationale pour la Recherche et l'Enseignement en Neurosciences (NINCDS-AIREN) [29] and the Hachinski Ischemic Score [30]. Additionally, 11 subjects were healthy controls (HC), 8 suffered from DLB, 3

from AD+VaD, 3 from PD, 1 from CBS, and 3 had ND (Table 1). All participants were neurologically examined. Complete blood tests including determination of vitamin B12 and folic acid (B9) levels, thyroid function, serology for Lyme's disease and syphilis were obtained for each patient. They were also tested neuropsychologically using the Alzheimer's Disease Assessment Scale-cognitive subscale (ADAS-Cog), Mini-Mental State Examination (MMSE), and Montreal Cognitive Assessment (MoCA). Informed consent for participation was obtained from all patients and HC, and all procedures were approved by the Central Ethical Committee of the University of Zagreb Medical School (case no. 380-59-10106-18-111/126, class 641-01/18-02/01 from June 20, 2018) and Ethical Committee of the Clinical Hospital Center Zagreb (case no. 02/21 AG, class 8.1-18/82-2 from April 24, 2018) and done in accord with the Helsinki Declaration [31].

2.2. DNA analysis

Genomic DNA was extracted from peripheral blood using the salting-out method [32]. *MAOB* rs1799836 and *APOE* rs7412 and rs429358 (for *APOE* variants $\epsilon 2$, $\epsilon 3$, and $\epsilon 4$) SNPs were determined using primers and probes purchased from Applied Biosystems as TaqMan® SNP Genotyping Assays (C_8878790_10 for rs1799836, C_904973_10 for rs7412 and C_3084793_20 for rs429358) on an ABI Prism 7300 Real Time PCR System apparatus (Applied Biosystems). *APOE* $\epsilon 2$ variant is determined by both rs429358 and rs7412 T allele, the $\epsilon 4$ variant is determined by both rs429358 and rs7412 C allele, while $\epsilon 3$ variant is determined by rs429358 T allele and rs7412 C allele. All genotyping procedures were done by a researcher who was blind to all clinical data according to the procedures described by Applied Biosystems. Out of 253, 74 samples (29%) were genotyped again as a quality control for genotyping analyses.

2.3. Statistical analysis

The frequencies of *APOE* genotypes and alleles among subjects with different *MAOB* rs1799836 genotypes and alleles was analyzed using a χ^2 -test. A correction for

pairwise comparisons was applied. Correction for sex was done using binary and multinomial logistic regression analysis. All statistical analyses were done using SPSS 19.0.1 (SPSS, Chicago, IL, USA) with the level of statistical significance set at $\alpha = 0.05$.

3. RESULTS

There was a significant increase in the frequency of *APOE* $\epsilon 4/\epsilon 4$ homozygotes among patients carrying the AA *MAOB* rs1799836 genotype in comparison to patients carrying other *MAOB* rs1799836 genotypes [in the group of AD patients ($\chi^2=12.815$; $df=4$; $p=0.012$), in the group of AD and MCI patients and HC ($\chi^2=14.081$; $df=4$; $p=0.007$), and in the group of all subjects ($\chi^2=16.316$; $df=4$; $p=0.003$)] (**Figure 1**) or G allele [in the group of AD and MCI patients ($\chi^2=11.509$; $df=2$; $p=0.003$), in the group of AD and MCI patients and HC ($\chi^2=13.368$; $df=2$; $p=0.001$), and in the group of all subjects ($\chi^2=15.541$; $df=2$; $p<0.001$)] (**Figure 1**; **Figure 2**). Also, frequency of *APOE* $\epsilon x/\epsilon x$ genotype ($x = 2$ or 3) (**Figure 1E-G**) and *APOE* $\epsilon 3/\epsilon 3$ genotype (**Figure 2**) is increased among carriers of G allele in *MAOB* rs1799836 polymorphism. There was significantly higher frequency of *APOE* $\epsilon 4$ carriers ($\epsilon 4/\epsilon 4 + \epsilon 4/\epsilon x$ genotypes) among AA *MAOB* rs1799836 homozygotes [**Figures 3A-C**; in the group of AD and MCI patients ($\chi^2=8.076$; $df=2$; $p=0.018$), in the group of AD and MCI patients and HC ($\chi^2=10.086$; $df=2$; $p=0.006$), and in the group of all subjects ($\chi^2=9.828$; $df=2$; $p=0.007$), **Figures 3D-G**; in the group of AD patients ($\chi^2=6.835$; $df=1$; $p=0.009$), in the group of AD and MCI patients ($\chi^2=7.999$; $df=1$; $p=0.005$), in the group of AD and MCI patients and HC ($\chi^2=9.849$; $df=1$; $p=0.002$), and in the group of all subjects ($\chi^2=9.551$; $df=1$; $p=0.002$)]. Also, significantly higher frequency of *APOE* $\epsilon 4$ non-carriers was observed among carriers of G allele in *MAOB* rs1799836 polymorphism (**Figures 3D-G**).

Since *MAOB* gene is located on X chromosome, it is not surprising that we observed statistically significant difference in distribution of *MAOB* rs1799836 genotypes between males and females ($\chi^2=69.974$; $df=2$; $p<0.001$). Thus, we tested distribution of *MAOB* rs1799836 genotypes between patients with different *APOE* genotypes but with

MAOB rs1799836 genotypes being adjusted for sex (**Table 2**). Logistic regression model revealed that AA *MAOB* rs1799836 genotype had significant association with *APOE* $\epsilon 4$ allele ($p=0.002$) (**Table 2**), *APOE* $\epsilon 4/\epsilon 4$ ($p=0.022$) and *APOE* $\epsilon 4/\epsilon x$ genotype ($p=0.015$) even when adjusted for sex (**Table 3**). No significant association between sex and *APOE* genotype was detected ($p=0.781$) (**Table 2**).

We also tested if *MAOB* and *APOE* polymorphisms are good predictors of AD using logistic regression. Neither *MAOB* nor *APOE* were proved as good predictors of AD (**Supplementary table 1**). However, possible cause for these results is because we included only 11 HCs in this study. Thus, these results should be verified on the bigger cohort.

4. DISCUSSION

The goal of this study was to test whether *MAOB* rs1799836 polymorphism is associated with *APOE* genotype. This polymorphism affects *MAOB* transcription and consequently influences the amount of the produced protein [23]. We showed that the frequency of *APOE* $\epsilon 4/\epsilon 4$ homozygotes and *APOE* $\epsilon 4$ carriers was significantly increased among patients carrying AA *MAOB* rs1799836 genotype.

The dopaminergic system is affected in AD and in animal models of AD [33–36]. The degeneration of main dopaminergic nuclei, the ventral tegmental area (VTA) [37] and the substantia nigra pars compacta [38] has also been observed in animal models of AD. Altered connectivity between the VTA and other brain regions implicated in AD pathology have been observed [39], further supporting dopaminergic system dysregulation in AD. It was even suggested that VTA volume could represent early neuroimaging biomarker of neurodegeneration [40,41]. Krashia et al. suggested that dopaminergic neurons in VTA could be more prone to cell death than other neuronal cells since they have long unmyelinated axons that innervate cortex and also since due to their autonomous pacemaker activity [self-generated activity that maintains dopamine levels in the brain [42]] they require high levels of energy and efficient mitochondrial function [36]. Levels of dopamine, dopamine metabolites, activity of dopamine β -

hydroxylase (DBH), expression and availability of dopamine receptors are decreased in AD [19–21,43–47]. Additionally, polymorphisms in genes encoding proteins and enzymes of the dopaminergic system are associated with behavioral and psychological symptoms of dementia observed in early AD [48,49].

Increased activity of MAOB has been reported in AD [15–18]. MAOB activity is influenced by various medications, smoking, ethnicity, gender, and ageing [50–57]. Thus, it was suggested that MAOB represents a molecular link between AD pathogenesis and lifestyle [58]. It was proposed that MAOB might serve as peripheral biomarker of AD in view of high levels of sensitivity and specificity in differentiating AD patients and HC [58,59]. Also, neuroimaging study of Rodriguez-Vieitez et al. showed potential of MAOB as an early biomarker of AD since binding of MAOB ligand (11C-deuterium-l-deprenyl) was increased in presymptomatic early-onset familial AD cases [60]. MAOB expression is increased in the neocortex and hippocampus in postmortem AD brains [61–63]. An increase in MAOB expression was detected in reactive astrocytes around amyloid plaques [64]. Such increases in expression or activity result in higher production of H₂O₂ — a by-product of MAOB activity — and of reactive oxygen species-induced oxidative stress [65]. Increased MAOB activity also contributes to mitochondrial dysfunction in AD [66]. Additionally, Schedin-Weiss et al. showed that MAOB is γ -secretase associated protein that can regulate levels of A β ₁₋₄₂. They proved that MAOB silencing by siRNA reduces intraneuronal A β ₁₋₄₂ levels, while MAOB overexpression leads to increase in A β ₁₋₄₂ levels [18]. MAOB inhibitors were in fact tested as potential therapeutics in AD (selegiline, lazabemide, sembragiline) with purpose to reduce cognitive decline [67–69]. Additionally, several multi-target drugs have been designed to inhibit MAOB in AD. For example, ASS234 inhibits both monoamine oxidases (MAO-A/MAO-B) and cholinesterases [70] as do PF1901N [71], M30D [72] and ladostigil [73]. Since AD is product of various factors, such as age, genetic predisposition, lifestyle and environmental risk factors, it is not surprising that in addition to age and lifestyle factors that affect MAOB activity, some genetic factors could also

change enzyme's activity. It was reported that *MAOB* rs1799836 polymorphism can affect *MAOB* transcription and translation. Thus, this polymorphism could affect enzyme's activity that could lead to altered concentration of dopamine in synapses [23]. However, conflicting results were obtained in the studies with both A [74] and G [75] allele in *MAOB* rs1799836 polymorphism being associated with increased MAOB activity. Possible cause of discrepancies between these studies is that Balciuniene et al. [74] measured MAOB activity in the brain, while Garpenstrand et al. [75] measured platelet MAOB activity. Additional two studies that measured platelet MAOB activity [76] and dopamine turnover in the brain [77] observed the association of A allele in *MAOB* rs1799836 polymorphism with increased MAOB activity. However, some studies failed to confirm the association of *MAOB* rs1799836 polymorphism and MAOB activity [53,56,57,78]. There is a little information in the literature on the interaction between *APOE* and MAOB. However, Veitinger et al. reached high levels of specificity and sensitivity in differentiating AD patients and HC when combining platelet MAOB activity and *APOE* ϵ 4 allele [58]. Also, recent study of Quartey et al. showed that MAOB activity was increased in hippocampus and cortex of AD donors who carried *APOE* ϵ 4 allele in comparison to *APOE* ϵ 4 non-carriers. They also proved that *in vitro* overexpression of human *APOE4* in C6 and in HT-22 cell cultures increases MAOB activity. Since MAOB activity was increased in C6 and in HT-22 cells without changes in MAOB protein levels, authors suggested that *APOE4* might through post-translational mechanisms influence MAOB function [63]. Since the results of our study showed that the frequency of *APOE* ϵ 4/ ϵ 4 homozygotes and *APOE* ϵ 4 carriers was significantly increased among patients carrying AA *MAOB* rs1799836 genotype (and by taking into account conflicting results of previous studies on the influence of *MAOB* rs1799836 polymorphism on MAOB activity), it should be further validated in the cohort of AD patients if *MAOB* rs1799836 polymorphism affects MAOB activity.

CONCLUSION

The present study provides more support for *MAOB* rs1799836 polymorphism as a potential genetic biomarker of

AD. We previously reported an association of this polymorphism with $A\beta_{1-42}$ measured in CSF [24], and now we also showed its association with *APOE* $\epsilon 4$ genotype. By taking these studies into account, we suggest that patients carrying the AA *MAOB* rs1799836 genotype could have a higher risk for AD development. However, further longitudinal studies should test if MCI patients carrying AA *MAOB* rs1799836 genotype would actually develop AD. Also, further studies should investigate the distribution of *MAOB* rs1799836 genotypes between AD, MCI patients and HC and also the association of this polymorphism with neuroimaging biomarkers of AD. In conclusion, the present results together with our previous observations [24] indicate that the *MAOB* rs1799836 polymorphism could be an important genetic biomarker of AD.

LIST OF ABBREVIATIONS

$A\beta$, amyloid β protein; AD, Alzheimer's disease; ADAS-Cog, Alzheimer's Disease Assessment Scale-cognitive subscale; APOE, apolipoprotein E; CBS, corticobasal syndrome; CSF, cerebrospinal fluid; DLB, dementia with Lewy bodies; FTD, frontotemporal dementia; HC, healthy control; HIS, Hachinski Ischemic Score; MAOB, monoamine oxidase B; MCI, mild cognitive impairment; MMSE, Mini-Mental State Examination; MoCA, Montreal Cognitive Assessment; ND, nonspecific dementia; NINCDS-AIREN, National Institute for Neurological Disorders and Stroke - Association Internationale pour la Recherche et l'Enseignement en Neurosciences; PD, Parkinson's disease; SNP, single nucleotide polymorphisms; VaD, vascular dementia; VTA, ventral tegmental area.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

This work was funded by The Croatian Science Foundation grant IP-2019-04-3584 ("Role of blood-brain barrier, innate immunity, and tau protein oligomerization in the

pathogenesis of Alzheimer's disease") to GŠ and by the Scientific Centre of Excellence for Basic, Clinical and Translational Neuroscience CoRE-NEURO ("Experimental and clinical research of hypoxic-ischemic damage in perinatal and adult brain"; GA KK01.1.1.01.0007 funded by the European Union through the European Regional Development Fund), and in part by the NIH grant P50 AG005138 to PRH.

REFERENCES

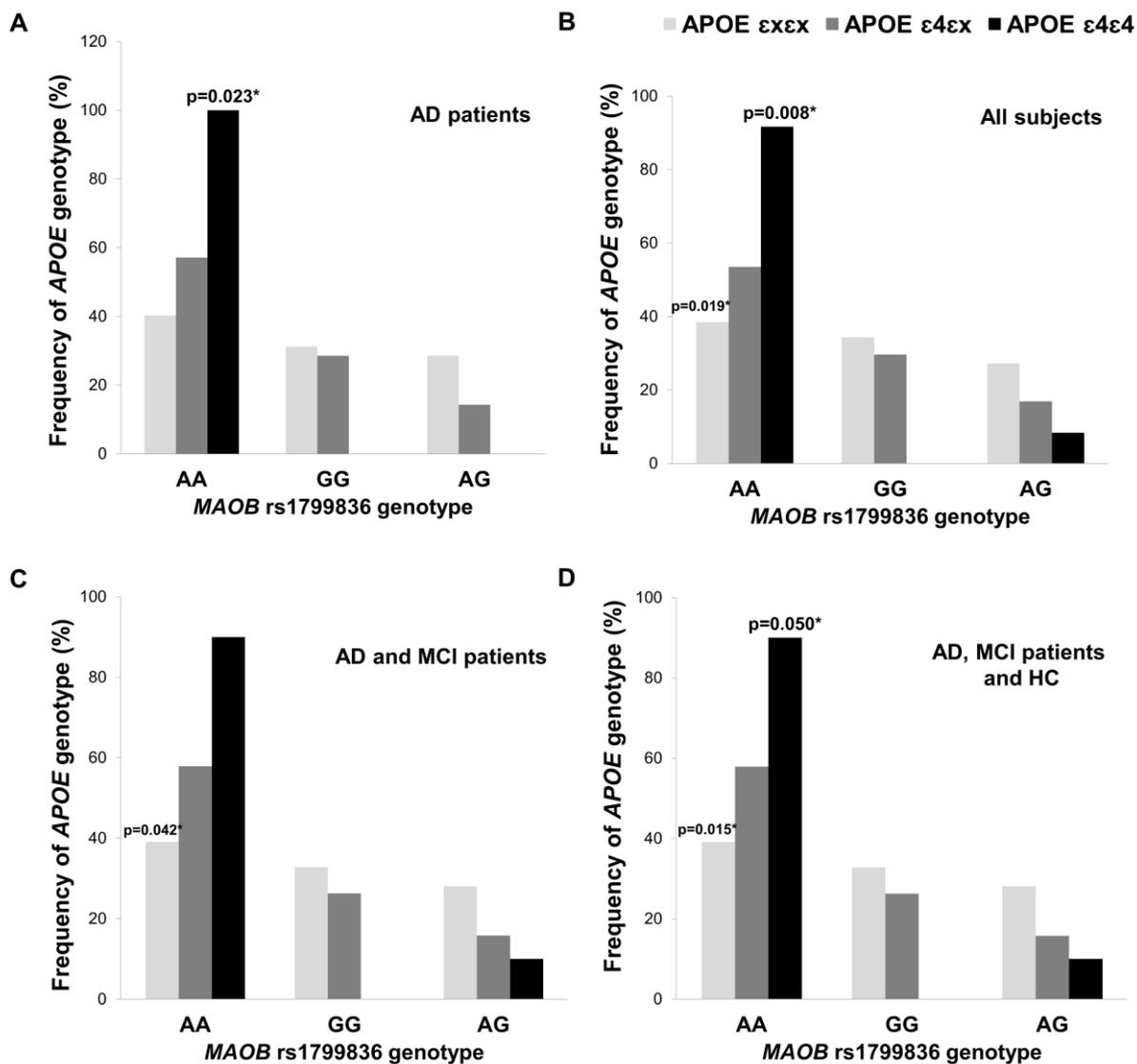
1. Strittmatter, W.J.; Saunders, A.M.; Schmechel, D.; Pericak-Vance, M.; Enghild, J.; Salvesen, G.S.; Roses, A.D. Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc. Natl. Acad. Sci. U. S. A.* **1993**, *90*, 1977–1981, doi:10.1073/pnas.90.5.1977.
2. Saunders, A.M.; Strittmatter, W.J.; Schmechel, D.; George-Hyslop, P.H.; Pericak-Vance, M.A.; Joo, S.H.; Rosi, B.L.; Gusella, J.F.; Crapper-MacLachlan, D.R.; Alberts, M.J. Association of apolipoprotein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. *Neurology* **1993**, *43*, 1467–1472, doi:10.1212/WNL.43.8.1467.
3. Bu, G. Apolipoprotein E and its receptors in Alzheimer's disease: pathways, pathogenesis and therapy. *Nat. Rev. Neurosci.* **2009**, *10*, 333–344, doi:10.1038/nrn2620.
4. Belbin, O.; Dunn, J.L.; Ling, Y.; Morgan, L.; Chappell, S.; Beaumont, H.; Warden, D.; Smith, D.A.; Kalsheker, N.; Morgan, K. Regulatory region single nucleotide polymorphisms of the apolipoprotein E gene and the rate of cognitive decline in Alzheimer's disease. *Hum. Mol. Genet.* **2007**, *16*, 2199–2208, doi:10.1093/hmg/ddm171.
5. Conejero-Goldberg, C.; Gomar, J.J.; Bobes-Bascaran, T.; Hyde, T.M.; Kleinman, J.E.; Herman, M.M.; Chen, S.; Davies, P.; Goldberg, T.E. APOE2 enhances neuroprotection against Alzheimer's disease through multiple molecular mechanisms. *Mol. Psychiatry* **2014**, *19*, 1243–1250, doi:10.1038/mp.2013.194.
6. Strittmatter, W.J. Medicine. Old drug, new hope for Alzheimer's disease. *Science (80-.)*. **2012**, *335*, 1447–1448, doi:10.1126/science.1220725.
7. Kunkle, B.W.; Grenier-Boley, B.; Sims, R.; Bis, J.C.; Damotte, V.; Naj, A.C.; Boland, A.; Vronskaya, M.; van der Lee, S.J.; Amlie-Wolf, A.; et al. Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates $A\beta$, tau, immunity and lipid processing. *Nat. Genet.* **2019**, *51*, 414–430, doi:10.1038/s41588-019-0358-2.

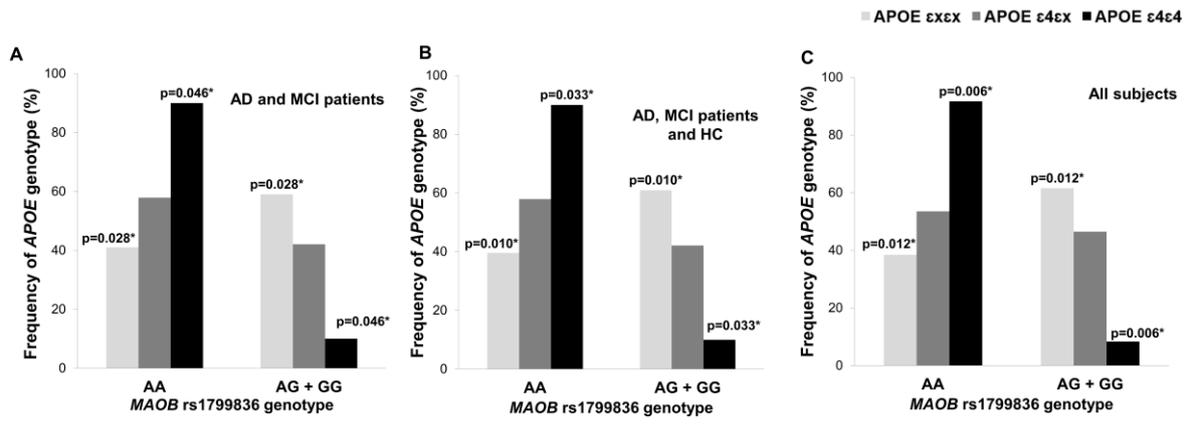
8. Naj, A.C.; Jun, G.; Beecham, G.W.; Wang, L.-S.; Vardarajan, B.N.; Buross, J.; Gallins, P.J.; Buxbaum, J.D.; Jarvik, G.P.; Crane, P.K.; et al. Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease. *Nat. Genet.* **2011**, *43*, 436–441, doi:10.1038/ng.801.
9. Hollingworth, P.; Harold, D.; Sims, R.; Gerrish, A.; Lambert, J.-C.; Carrasquillo, M.M.; Abraham, R.; Hamshere, M.L.; Pahwa, J.S.; Moskvin, V.; et al. Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. *Nat. Genet.* **2011**, *43*, 429–435, doi:10.1038/ng.803.
10. Harold, D.; Abraham, R.; Hollingworth, P.; Sims, R.; Gerrish, A.; Hamshere, M.L.; Pahwa, J.S.; Moskvin, V.; Dowzell, K.; Williams, A.; et al. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat. Genet.* **2009**, *41*, 1088–1093, doi:10.1038/ng.440.
11. Lambert, J.-C.; Heath, S.; Even, G.; Campion, D.; Sleegers, K.; Hiltunen, M.; Combarros, O.; Zelenika, D.; Bullido, M.J.; Tavernier, B.; et al. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat. Genet.* **2009**, *41*, 1094–1099, doi:10.1038/ng.439.
12. Deming, Y.; Li, Z.; Kapoor, M.; Harari, O.; Del-Aguila, J.L.; Black, K.; Carrell, D.; Cai, Y.; Fernandez, M.V.; Budde, J.; et al. Genome-wide association study identifies four novel loci associated with Alzheimer's endophenotypes and disease modifiers. *Acta Neuropathol.* **2017**, *133*, 839–856, doi:10.1007/s00401-017-1685-y.
13. Cruchaga, C.; Karch, C.M.; Jin, S.C.; Benitez, B.A.; Cai, Y.; Guerreiro, R.; Harari, O.; Norton, J.; Budde, J.; Bertelsen, S.; et al. Rare coding variants in the phospholipase D3 gene confer risk for Alzheimer's disease. *Nature* **2014**, *505*, 550–554, doi:10.1038/nature12825.
14. Jonsson, T.; Stefansson, H.; Steinberg, S.; Jonsdottir, I.; Jonsson, P. V.; Snaedal, J.; Bjornsson, S.; Huttenlocher, J.; Levey, A.I.; Lah, J.J.; et al. Variant of TREM2 associated with the risk of Alzheimer's disease. *N. Engl. J. Med.* **2013**, *368*, 107–116, doi:10.1056/NEJMoa1211103.
15. Adolfsson, R.; Gottfries, C.-G.; Oreland, L.; Wiberg, Å.; Winblad, B. Increased activity of brain and platelet monoamine oxidase in dementia of Alzheimer type. *Life Sci.* **1980**, *27*, 1029–1034, doi:10.1016/0024-3205(80)90025-9.
16. Kennedy, B.P.; Ziegler, M.G.; Alford, M.; Hansen, L.A.; Thal, L.J.; Masliah, E. Early and persistent alterations in prefrontal cortex MAO A and B in Alzheimer's disease. *J. Neural Transm.* **2003**, *110*, 789–801, doi:10.1007/s00702-003-0828-6.
17. Zellner, M.; Baureder, M.; Rappold, E.; Bugert, P.; Kotzailias, N.; Babeluk, R.; Baumgartner, R.; Attems, J.; Gerner, C.; Jellinger, K.; et al. Comparative platelet proteome analysis reveals an increase of monoamine oxidase-B protein expression in Alzheimer's disease but not in non-demented Parkinson's disease patients. *J. Proteomics* **2012**, *75*, 2080–2092, doi:10.1016/j.jprot.2012.01.014.
18. Schedin-Weiss, S.; Inoue, M.; Hromadkova, L.; Teranishi, Y.; Yamamoto, N.G.; Wichager, B.; Bogdanovic, N.; Winblad, B.; Sandebring-Matton, A.; Frykman, S.; et al. Monoamine oxidase B is elevated in Alzheimer disease neurons, is associated with γ -secretase and regulates neuronal amyloid β -peptide levels. *Alzheimers. Res. Ther.* **2017**, *9*, 57, doi:10.1186/s13195-017-0279-1.
19. Tohgi, H.; Ueno, M.; Abe, T.; Takahashi, S.; Nozaki, Y. Concentrations of monoamines and their metabolites in the cerebrospinal fluid from patients with senile dementia of the Alzheimer type and vascular dementia of the Binswanger type. *J. Neural Transm. Park. Dis. Dement. Sect.* **1992**, *4*, 69–77, doi:10.1007/bf02257623.
20. Pinessi, L.; Rainero, I.; De Gennaro, T.; Gentile, S.; Portaleone, P.; Bergamasco, B. Biogenic amines in cerebrospinal fluid and plasma of patients with dementia of Alzheimer type. *Funct. Neurol.* **1987**, *2*, 51–58.
21. Sjögren, M.; Minthon, L.; Passant, U.; Blennow, K.; Wallin, A. Decreased monoamine metabolites in frontotemporal dementia and Alzheimer's disease. *Neurobiol. Aging* **1998**, *19*, 379–384, doi:10.1016/s0197-4580(98)00086-4.
22. Finberg, J.P.M.; Rabey, J.M. Inhibitors of MAO-A and MAO-B in psychiatry and neurology. *Front. Pharmacol.* **2016**, *7*, 340, doi:10.3389/fphar.2016.00340.
23. Jakubauskiene, E.; Janaviciute, V.; Peculiene, I.; Söderkvist, P.; Kanopka, A. G/A polymorphism in intronic sequence affects the processing of MAO-B gene in patients with Parkinson disease. *FEBS Lett.* **2012**, *586*, 3698–3704, doi:10.1016/j.febslet.2012.08.028.
24. Babić Leko, M.; Nikolac Perković, M.; Klepac, N.; Švob Štrac, D.; Borovečki, F.; Pivac, N.; Hof, P.R.; Šimić, G. Relationships of Cerebrospinal Fluid Alzheimer's Disease Biomarkers and COMT, DBH, and MAOB Single Nucleotide Polymorphisms. *J. Alzheimer's Dis.* **2020**, *73*, 135–145, doi:10.3233/JAD-190991.
25. McKhann, G.M.; Knopman, D.S.; Chertkow, H.; Hyman, B.T.; Jack, C.R.; Kawas, C.H.; Klunk, W.E.; Koroshetz, W.J.; Manly, J.J.; Mayeux, R.; et al. The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on

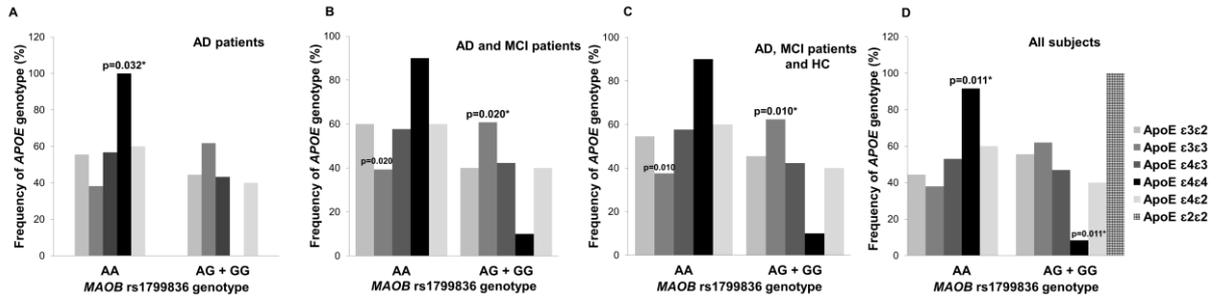
- diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement.* **2011**, *7*, 263–269, doi:10.1016/j.jalz.2011.03.005.
26. Petersen, R.C.; Smith, G.E.; Waring, S.C.; Ivnik, R.J.; Tangalos, E.G.; Kokmen, E. Mild cognitive impairment: clinical characterization and outcome. *Arch. Neurol.* **1999**, *56*, 303–308, doi:10.1001/archneur.56.3.303.
 27. Albert, M.S.; DeKosky, S.T.; Dickson, D.; Dubois, B.; Feldman, H.H.; Fox, N.C.; Gamst, A.; Holtzman, D.M.; Jagust, W.J.; Petersen, R.C.; et al. The diagnosis of mild cognitive impairment due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement.* **2011**, *7*, 270–279, doi:10.1016/j.jalz.2011.03.008.
 28. Neary, D.; Snowden, J.S.; Gustafson, L.; Passant, U.; Stuss, D.; Black, S.; Freedman, M.; Kertesz, A.; Robert, P.H.; Albert, M.; et al. Frontotemporal lobar degeneration: a consensus on clinical diagnostic criteria. *Neurology* **1998**, *51*, 1546–1554, doi:10.1212/wnl.51.6.1546.
 29. Román, G.C.; Tatemichi, T.K.; Erkinjuntti, T.; Cummings, J.L.; Masdeu, J.C.; Garcia, J.H.; Amaducci, L.; Orgogozo, J.M.; Brun, A.; Hofman, A. Vascular dementia: diagnostic criteria for research studies. Report of the NINDS-AIREN International Workshop. *Neurology* **1993**, *43*, 250–260, doi:10.1212/wnl.43.2.250.
 30. Hachinski, V.C.; Iliff, L.D.; Zilhka, E.; Du Boulay, G.H.; McAllister, V.L.; Marshall, J.; Russell, R.W.; Symon, L. Cerebral blood flow in dementia. *Arch. Neurol.* **1975**, *32*, 632–637, doi:10.1001/archneur.1975.00490510088009.
 31. World Medical Association World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects. *JAMA* **2013**, *310*, 2191–2194, doi:10.1001/jama.2013.281053.
 32. Miller, S.A.; Dykes, D.D.; Polesky, H.F. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res.* **1988**, *16*, 1215, doi:10.1093/nar/16.3.1215.
 33. Šimić, G.; Babić Leko, M.; Wray, S.; Harrington, C.R.; Delalle, I.; Jovanov-Milošević, N.; Bažadona, D.; Buée, L.; de Silva, R.; Di Giovanni, G.; et al. Monoaminergic neuropathology in Alzheimer's disease. *Prog. Neurobiol.* **2017**, *151*, 101–138, doi:10.1016/j.pneurobio.2016.04.001.
 34. Babić Leko, M.; Hof, P.R.; Šimić, G. Alterations and interactions of subcortical modulatory systems in Alzheimer's disease. *Prog. Brain Res.* **2021**, *261*, 379–421, doi:10.1016/bs.pbr.2020.07.016.
 35. Trillo, L.; Das, D.; Hsieh, W.; Medina, B.; Moghadam, S.; Lin, B.; Dang, V.; Sanchez, M.M.; De Miguel, Z.; Ashford, J.W.; et al. Ascending monoaminergic systems alterations in Alzheimer's disease. Translating basic science into clinical care. *Neurosci. Biobehav. Rev.* **2013**, *37*, 1363–1379, doi:10.1016/j.neubiorev.2013.05.008.
 36. Krashia, P.; Nobili, A.; D'Amelio, M. Unifying Hypothesis of Dopamine Neuron Loss in Neurodegenerative Diseases: Focusing on Alzheimer's Disease. *Front. Mol. Neurosci.* **2019**, *12*, 123, doi:10.3389/fnmol.2019.00123.
 37. Nobili, A.; Latagliata, E.C.; Viscomi, M.T.; Cavallucci, V.; Cutuli, D.; Giacobuzzo, G.; Krashia, P.; Rizzo, F.R.; Marino, R.; Federici, M.; et al. Dopamine neuronal loss contributes to memory and reward dysfunction in a model of Alzheimer's disease. *Nat. Commun.* **2017**, *8*, 14727, doi:10.1038/ncomms14727.
 38. Chenyou Sun, C.; Xiaoming Ou, X.; Jerry M. Farley, J.M.; Craig Stockmeier, C.; Steven Bigler, S.; Roberta Diaz Brinton, R.D.; Jun Ming Wang, J.M. Allopregnanolone Increases the Number of Dopaminergic Neurons in Substantia Nigra of a Triple Transgenic Mouse Model of Alzheimer's Disease. *Curr. Alzheimer Res.* **2012**, *9*, 473–480, doi:10.2174/156720512800492567.
 39. Serra, L.; D'Amelio, M.; Di Domenico, C.; Dipasquale, O.; Marra, C.; Mercuri, N.B.; Caltagirone, C.; Cercignani, M.; Bozzali, M. In vivo mapping of brainstem nuclei functional connectivity disruption in Alzheimer's disease. *Neurobiol. Aging* **2018**, *72*, 72–82, doi:10.1016/j.neurobiolaging.2018.08.012.
 40. D'Amelio, M.; Serra, L.; Bozzali, M. Ventral Tegmental Area in Prodromal Alzheimer's Disease: Bridging the Gap between Mice and Humans. *J. Alzheimer's Dis.* **2018**, *63*, 181–183, doi:10.3233/JAD-180094.
 41. De Marco, M.; Venneri, A. Volume and Connectivity of the Ventral Tegmental Area are Linked to Neurocognitive Signatures of Alzheimer's Disease in Humans. *J. Alzheimer's Dis.* **2018**, *63*, 167–180, doi:10.3233/JAD-171018.
 42. Guzman, J.N.; Sánchez-Padilla, J.; Chan, C.S.; Surmeier, D.J. Robust Pacemaking in Substantia Nigra Dopaminergic Neurons. *J. Neurosci.* **2009**, *29*, 11011, doi:10.1523/JNEUROSCI.2519-09.2009.
 43. Zubenko, G.S.; Marquis, J.K.; Volicer, L.; Drenfeld, L.K.; Langlais, P.J.; Nixon, R.A. Cerebrospinal fluid levels of angiotensin- converting enzyme, acetylcholinesterase, and dopamine metabolites in dementia associated with Alzheimer's disease and Parkinson's disease: A correlative study. *Biol. Psychiatry* **1986**, *21*, 1365–1381, doi:10.1016/0006-3223(86)90328-8.
 44. Bareggi, S.R.; Franceschi, M.; Bonini, L.; Zecca, L.;

- Smirne, S. Decreased CSF Concentrations of Homovanillic Acid and γ -Aminobutyric Acid in Alzheimer's Disease. *Arch. Neurol.* **1982**, *39*, 709, doi:10.1001/archneur.1982.00510230035010.
45. Blennow, K.; Wallin, A.; Gottfries, C.G.; Lekman, A.; Karlsson, I.; Skoog, I.; Svennerholm, L. Significance of decreased lumbar CSF levels of HVA and 5-HIAA in Alzheimer's disease. *Neurobiol. Aging* **1992**, *13*, 107–113, doi:10.1016/0197-4580(92)90017-r.
46. Kempainen, N.; Laine, M.; Laakso, M.P.; Kaasinen, V.; Nagren, K.; Vahlberg, T.; Kurki, T.; Rinne, J.O. Hippocampal dopamine D2 receptors correlate with memory functions in Alzheimer's disease. *Eur. J. Neurosci.* **2003**, *18*, 149–154, doi:10.1046/j.1460-9568.2003.02716.x.
47. Kumar, U.; Patel, S.C. Immunohistochemical localization of dopamine receptor subtypes (D1R–D5R) in Alzheimer's disease brain. *Brain Res.* **2007**, *1131*, 187–196, doi:10.1016/j.brainres.2006.10.049.
48. Borroni, B.; Agosti, C.; Archetti, S.; Costanzi, C.; Bonomi, S.; Ghianda, D.; Lenzi, G.L.; Caimi, L.; Luca, M. Di; Padovani, A. Catechol-O-methyltransferase gene polymorphism is associated with risk of psychosis in Alzheimer Disease. *Neurosci. Lett.* **2004**, *370*, 127–129, doi:10.1016/j.neulet.2004.08.006.
49. Holmes, C.; Smith, H.; Ganderton, R.; Arranz, M.; Collier, D.; Powell, J.; Lovestone, S. Psychosis and aggression in Alzheimer's disease: the effect of dopamine receptor gene variation. *J. Neurol. Neurosurg. Psychiatry* **2001**, *71*, 777–779, doi:10.1136/jnnp.71.6.777.
50. Kiive, E.; Eensoo, D.; Harro, M.; Harro, J. Platelet monoamine oxidase activity in association with childhood aggressive and hyperactive behaviour: the effect of smoking? *Pers. Individ. Dif.* **2002**, *33*, 355–363, doi:10.1016/S0191-8869(01)00160-X.
51. Muck-Seler, D.; Sagud, M.; Mustapic, M.; Nedic, G.; Babic, A.; Mihaljevic Peles, A.; Jakovljevic, M.; Pivac, N. The effect of lamotrigine on platelet monoamine oxidase type B activity in patients with bipolar depression. *Prog. Neuro-Psychopharmacology Biol. Psychiatry* **2008**, *32*, 1195–1198, doi:10.1016/j.pnpbp.2008.03.004.
52. Muck-Seler, D.; Presecki, P.; Mimica, N.; Mustapic, M.; Pivac, N.; Babic, A.; Nedic, G.; Folnegovic-Smalc, V. Platelet serotonin concentration and monoamine oxidase type B activity in female patients in early, middle and late phase of Alzheimer's disease. *Prog. Neuro-Psychopharmacology Biol. Psychiatry* **2009**, *33*, 1226–1231, doi:10.1016/j.pnpbp.2009.07.004.
53. Nedic Erjavec, G.; Nenadic Sviglin, K.; Nikolac Perkovic, M.; Muck-Seler, D.; Jovanovic, T.; Pivac, N. Association of gene polymorphisms encoding dopaminergic system components and platelet MAO-B activity with alcohol dependence and alcohol dependence-related phenotypes. *Prog. Neuro-Psychopharmacology Biol. Psychiatry* **2014**, *54*, 321–327, doi:10.1016/j.pnpbp.2014.07.002.
54. Orelund, L. Platelet Monoamine Oxidase, Personality and Alcoholism: The Rise, Fall and Resurrection. *Neurotoxicology* **2004**, *25*, 79–89, doi:10.1016/S0161-813X(03)00115-3.
55. Pivac, N.; Mück-Šeler, D.; Šagud, M.; Jakovljević, M.; Mustapić, M.; Mihaljević-Peješ, A. Long-term sertraline treatment and peripheral biochemical markers in female depressed patients. *Prog. Neuro-Psychopharmacology Biol. Psychiatry* **2003**, *27*, 759–765, doi:10.1016/S0278-5846(03)00105-2.
56. Pivac, N.; Knezevic, J.; Mustapic, M.; Dezeljin, M.; Muck-Seler, D.; Kozaric-Kovacic, D.; Balija, M.; Matijevic, T.; Pavelic, J. The lack of association between monoamine oxidase (MAO) intron 13 polymorphism and platelet MAO-B activity among men. *Life Sci.* **2006**, *79*, 45–49, doi:10.1016/j.lfs.2005.12.030.
57. Pivac, N.; Knezevic, J.; Kozaric-Kovacic, D.; Dezeljin, M.; Mustapic, M.; Rak, D.; Matijevic, T.; Pavelic, J.; Muck-Seler, D. Monoamine oxidase (MAO) intron 13 polymorphism and platelet MAO-B activity in combat-related posttraumatic stress disorder. *J. Affect. Disord.* **2007**, *103*, 131–138, doi:10.1016/j.jad.2007.01.017.
58. Veitinger, M.; Oehler, R.; Umlauf, E.; Baumgartner, R.; Schmidt, G.; Gerner, C.; Babeluk, R.; Attems, J.; Mitulovic, G.; Rappold, E.; et al. A platelet protein biochip rapidly detects an Alzheimer's disease-specific phenotype. *Acta Neuropathol.* **2014**, *128*, 665–677, doi:10.1007/s00401-014-1341-8.
59. Veitinger, M.; Varga, B.; Guterres, S.B.; Zellner, M. Platelets, a reliable source for peripheral Alzheimer's disease biomarkers? *Acta Neuropathol. Commun.* **2014**, *2*, 65, doi:10.1186/2051-5960-2-65.
60. Rodriguez-Vieitez, E.; Saint-Aubert, L.; Carter, S.F.; Almkvist, O.; Farid, K.; Schöll, M.; Chiotis, K.; Thordardottir, S.; Graff, C.; Wall, A.; et al. Diverging longitudinal changes in astrocytosis and amyloid PET in autosomal dominant Alzheimer's disease. *Brain* **2016**, *139*, 922–936, doi:10.1093/brain/awv404.
61. Reinikainen, K.J.; Paljärvi, L.; Halonen, T.; Malminen, O.; Kosma, V.M.; Laakso, M.; Riekkinen, P.J. Dopaminergic system and monoamine oxidase-B activity in Alzheimer's disease. *Neurobiol. Aging* **1988**, *9*, 245–252, doi:10.1016/s0197-4580(88)80061-7.
62. Emilsson, L.; Sætre, P.; Balciuniene, J.; Castensson, A.; Cairns, N.; Jazin, E.E. Increased monoamine oxidase messenger RNA expression levels in frontal cortex of Alzheimer's disease patients. *Neurosci.*

- Lett.* **2002**, 326, 56–60, doi:10.1016/s0304-3940(02)00307-5.
63. Quartey, M.O.; Nyarko, J.N.K.; Pennington, P.R.; Heistad, R.M.; Klassen, P.C.; Baker, G.B.; Mousseau, D.D. Alzheimer Disease and Selected Risk Factors Disrupt a Co-regulation of Monoamine Oxidase-A/B in the Hippocampus, but Not in the Cortex. *Front. Neurosci.* **2018**, 12, doi:10.3389/FNINS.2018.00419.
 64. Saura, J.; Luque, J.M.; Cesura, A.M.; Prada, M. Da; Chan-Palay, V.; Huber, G.; Löffler, J.; Richards, J.G. Increased monoamine oxidase b activity in plaque-associated astrocytes of Alzheimer brains revealed by quantitative enzyme radioautography. *Neuroscience* **1994**, 62, 15–30, doi:10.1016/0306-4522(94)90311-5.
 65. Riederer, P.; Danielczyk, W.; Grünblatt, E. Monoamine Oxidase-B Inhibition in Alzheimer's Disease. *Neurotoxicology* **2004**, 25, 271–277, doi:10.1016/S0161-813X(03)00106-2.
 66. Wong, K.Y.; Roy, J.; Fung, M.L.; Heng, B.C.; Zhang, C.; Lim, L.W. Relationships between mitochondrial dysfunction and neurotransmission failure in Alzheimer's disease. *Aging Dis.* **2020**, 11, 1291–1316, doi:10.14336/AD.2019.1125.
 67. Sano, M.; Ernesto, C.; Thomas, R.G.; Klauber, M.R.; Schafer, K.; Grundman, M.; Woodbury, P.; Growdon, J.; Cotman, C.W.; Pfeiffer, E.; et al. A Controlled Trial of Selegiline, Alpha-Tocopherol, or Both as Treatment for Alzheimer's Disease. *N. Engl. J. Med.* **1997**, 336, 1216–1222, doi:10.1056/NEJM199704243361704.
 68. Magni, G.; Meibach, R.C. Lazabemide for the long-term treatment of Alzheimer's disease. *Eur. Neuropsychopharmacol.* **1999**, 9, 142, doi:10.1016/S0924-977X(99)80017-0.
 69. Nave, S.; Doody, R.S.; Boada, M.; Grimmer, T.; Savola, J.-M.; Delmar, P.; Pauly-Evers, M.; Nikolcheva, T.; Czech, C.; Borroni, E.; et al. Sembragiline in Moderate Alzheimer's Disease: Results of a Randomized, Double-Blind, Placebo-Controlled Phase II Trial (MAYflOwer RoAD). *J. Alzheimer's Dis.* **2017**, 58, 1217–1228, doi:10.3233/JAD-161309.
 70. Marco-Contelles, J.; Unzeta, M.; Bolea, I.; Esteban, G.; Ramsay, R.R.; Romero, A.; Martínez-Murillo, R.; Carreiras, M.C.; Ismaili, L. ASS234, As a New Multi-Target Directed Propargylamine for Alzheimer's Disease Therapy. *Front. Neurosci.* **2016**, 10, 294, doi:10.3389/fnins.2016.00294.
 71. Bolea, I.; Colivicchi, M.A.; Ballini, C.; Marco-Contelles, J.; Tipton, K.F.; Unzeta, M.; Della Corte, L. Neuroprotective Effects of the MAO-B Inhibitor, PF9601N, in an in vivo Model of Excitotoxicity. *CNS Neurosci. Ther.* **2014**, 20, 641–650, doi:10.1111/cns.12271.
 72. Zheng, H.; Youdim, M.B.H.; Fridkin, M. Site-Activated Chelators Targeting Acetylcholinesterase and Monoamine Oxidase for Alzheimer's Therapy. *ACS Chem. Biol.* **2010**, 5, 603–610, doi:10.1021/cb900264w.
 73. Schneider, L.S.; Geffen, Y.; Rabinowitz, J.; Thomas, R.G.; Schmidt, R.; Ropele, S.; Weinstock, M.; Ladostigil Study Group Low-dose ladostigil for mild cognitive impairment. *Neurology* **2019**, 93, e1474–e1484, doi:10.1212/WNL.0000000000008239.
 74. Balciuniene, J.; Emilsson, L.; Orelund, L.; Pettersson, U.; Jazin, E. Investigation of the functional effect of monoamine oxidase polymorphisms in human brain. *Hum. Genet.* **2002**, 110, 1–7, doi:10.1007/s00439-001-0652-8.
 75. Garpenstrand, H.; Ekblom, J.; Forslund, K.; Rylander, G.; Orelund, L. Platelet monoamine oxidase activity is related to MAOB intron 13 genotype. *J. Neural Transm.* **2000**, 107, 523–530, doi:10.1007/s007020070075.
 76. Kakinuma, S.; Beppu, M.; Sawai, S.; Nakayama, A.; Hirano, S.; Yamanaka, Y.; Yamamoto, T.; Masafumi, C.; Aisihær, X.; Aersilan, A.; et al. Monoamine oxidase B rs1799836 G allele polymorphism is a risk factor for early development of levodopa-induced dyskinesia in Parkinson's disease. *eNeurologicalSci* **2020**, 19, 100239, doi:10.1016/j.ensci.2020.100239.
 77. Löhle, M.; Mangone, G.; Wolz, M.; Beuthien-Baumann, B.; Oehme, L.; van den Hoff, J.; Kotzerke, J.; Reichmann, H.; Corvol, J.C.; Storch, A. Functional monoamine oxidase B gene intron 13 polymorphism predicts putaminal dopamine turnover in de novo Parkinson's disease. *Mov. Disord.* **2018**, 33, 1496–1501, doi:10.1002/mds.27466.
 78. Tunbridge, E.M.; Narajos, M.; Harrison, C.H.; Beresford, C.; Cipriani, A.; Harrison, P.J. Which Dopamine Polymorphisms Are Functional? Systematic Review and Meta-analysis of COMT, DAT, DBH, DDC, DRD1-5, MAOA, MAOB, TH, VMAT1, and VMAT2. *Biol. Psychiatry* **2019**, <https://doi.org/10.1016/j.biopsych.2019.05.014>, doi:10.1016/j.biopsych.2019.05.014.







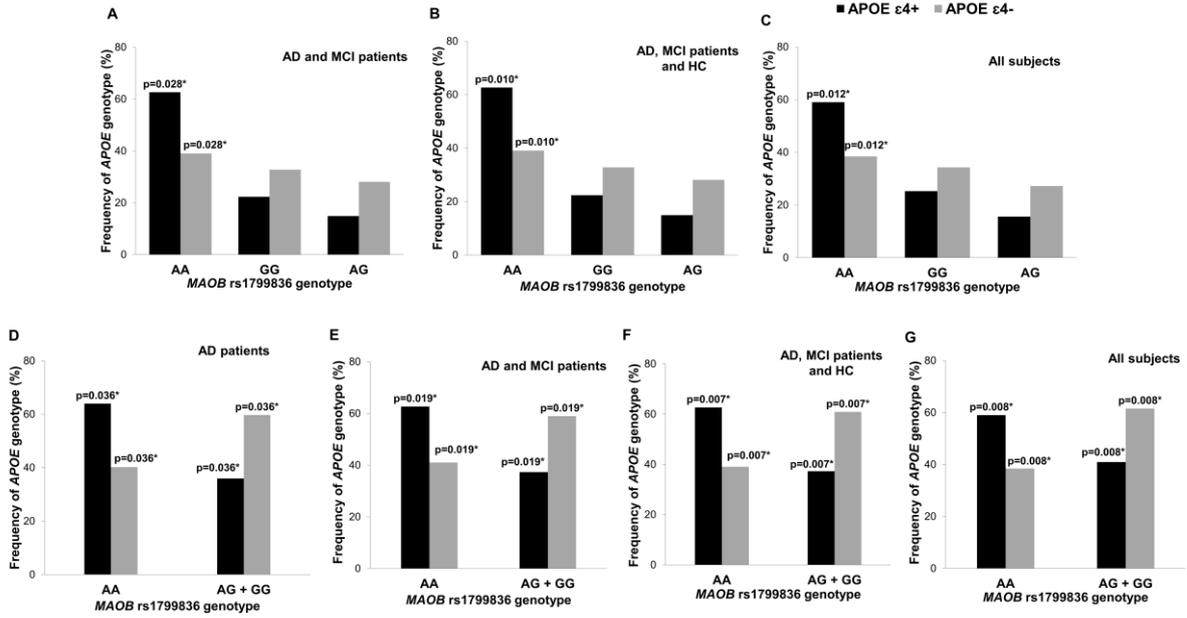


Table 1. Number of *MAOB* rs1799836 and *APOE* genotypes in AD and MCI patients, HC, and in patients with other causes of dementia.

	<i>MAOB</i>			<i>APOE</i>						Age	Sex	MMSE	Years of education
	AA	AG	GG	ε3ε2	ε3ε3	ε4ε3	ε4ε4	ε4ε2	ε2ε2	Median (25–75th percentile)	F/M	Mean ± SD	Median (25–75th percentile)
AD	63	28	36	9	68	37	8	5		73 (66-77)	68/59	19.9 ± 4.8	12 (8-14)
MCI	27	12	17	1	39	15	2			69 (58-74)	30/27	25.3 ± 3	12 (11-16)
HC	2	6	3	1	10					54 (45-61)	7/3	26.8 ± 2.5	12 (9-17)
VaD	7	5	3	2	8	4	1			72 (63-77)	6/7	23.1 ± 4.8	15 (9-16)
FTD	9	3	12	2	16	6	1			61 (56-66)	12/13	17.0 ± 5.7	12 (11-13)
DLB	3	2	3	1	5	2				71 (68-75)	3/5	20.4 ± 4.3	12 (4-16)
AD + VaD	2		1		2	1				78	0/3	19.3 ± 4.0	12
PD	1	1	1	1	1	1				65	1/2	15	8
CBS		1		1						51	1/0	24.0 ± 1.4	6
ND		1	2		2				1	68	2/1	20.7 ± 5.5	12

AD, Alzheimer's disease; AD + VaD, mixed dementia; APOE, apolipoprotein E; CBS, corticobasal syndrome; DLB, dementia with Lewy bodies; F, female; FTD, frontotemporal dementia; HC, healthy controls; M, male; MAOB, monoamine oxidase B; MCI, mild cognitive impairment; MMSE, Mini-Mental State Examination; ND, nonspecific dementia; PD, Parkinson's disease; SD, standard deviation; VaD, vascular dementia.

Table 2. Binary logistic regression analysis using *MAOB* rs1799836 genotype and *MAOB* genotype adjusted for sex as predictors of *APOE* genotype (carriers of $\epsilon 4$ allele vs $\epsilon 4$ non-carriers).

<i>APOE</i> $\epsilon 4$ allele								
Predictor	χ^2 , df, p	N	B	SE	Wald	p	OR	95% CI
Univariate model								
<i>MAOB</i> rs1799836 AA genotype	$\chi^2=9.518$, df=1, p=0.002	252	-0.835	0.274	9.329	0.002*	0.434	0.254-0.741
Sex	$\chi^2=0.078$, df=1, p=0.781	250	-0.075	0.270	0.077	0.781	0.928	0.546-1.575
Multivariate model								
<i>MAOB</i> rs1799836 AA genotype ^a	$\chi^2=9.638$, df=1, p=0.002	249	-0.846	0.275	9.453	0.002*	0.429	0.250-0.736

^aadjusted for sex. APOE, apolipoprotein E; MAOB, monoamine oxidase B. *p<0.05

Table 3. Multinomial logistic regression using *MAOB* rs1799836 genotype adjusted for sex as a predictor of *APOE* genotype ($\epsilon 4/\epsilon 4$, $\epsilon 4/\epsilon x$ and $\epsilon x/\epsilon x$ genotype [$x = 2$ or 3]).

Predictor	χ^2 , df, p	N	B	SE	Wald	p	OR	95% CI
<i>APOE</i> $\epsilon 4/\epsilon 4$ genotype								
<i>MAOB</i> rs1799836 AA genotype ^a	$\chi^2=21.501$, df=6, p=0.001	249	2.542	1.109	5.256	0.022*	12.709	1.446-111.695
<i>APOE</i> $\epsilon 4/\epsilon x$ genotype								
<i>MAOB</i> rs1799836 AA genotype ^a	$\chi^2=21.501$, df=6, p=0.001	249	1.056	0.432	5.961	0.015*	2.874	1.231-6.707

^aadjusted for sex. APOE, apolipoprotein E; MAOB, monoamine oxidase B. *p<0.05

Supplementary Table 1. Binary logistic regression analysis using *MAOB* rs1799836 genotype and *APOE* genotype as predictors of AD.

		Diagnosis – AD vs HC									
No.	Predictor	χ^2 , df, p	N	B	SE	Wald	p	OR	95% CI		
Univariate models											
1.	<i>MAOB</i> rs1799836 genotype	AA	138			5.403	0.067				
		GG				-1.910	0.847	5.077	0.024*	0.148	0.028-0.780
		AG				-0.944	0.751	1.583	0.208	0.389	0.089-1.694
2.	<i>MAOB</i> rs1799836 AA vs AG + GG genotype		138			3.447	0.063	0.226	0.047-1.086		
3.	<i>APOE</i> genotype	$\epsilon\chi\epsilon\chi$	138			0.000	1.000				
		$\epsilon 4\epsilon\chi$				19.257	14210.4	0.000	0.999	230782167	
		$\epsilon 4\epsilon 4$				0.000	15504.8	0.000	1.000	1.000	
4.	<i>APOE</i> genotype $\epsilon 4+$ vs $\epsilon 4-$		138			5684.14	0.000	0.997	0.000		
Multivariate models											
1.	<i>MAOB</i> rs1799836 genotype	AA	138			3.051	0.218				
		GG				-1.442	0.863	2.792	0.095	0.237	0.044-1.283
		AG				-0.780	0.766	1.037	0.309	0.458	0.102-2.058
	<i>APOE</i> genotype	$\epsilon\chi\epsilon\chi$				0.000	1.000				
		$\epsilon 4\epsilon\chi$	18.462	14210.37	0.000	0.999	104224200				
		$\epsilon 4\epsilon 4$	-0.510	15459.19	0.000	1.000	0.600				
2.	<i>MAOB</i> rs1799836 AA vs AG + GG genotype		138			1.851	0.174	0.330	0.067-1.631		
	<i>APOE</i> genotype $\epsilon 4+$ vs $\epsilon 4-$					5584.8	0.000	0.997	0.000		

AD, Alzheimer's disease; APOE, apolipoprotein E; HC, healthy controls; MAOB, monoamine oxidase B; No, number of models. *p<0.05