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Duraković, Nadira; Krečak, Ivan; Perić, Zinaida; Milošević, Milan; Desnica, Lana; Pulanić, Dražen; Pusic, Iskra; Kušec, Vesna; Vrhovac, Radovan; Pavletic, Steven Z.; ...

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Glycoprotein YKL-40: a novel biomarker of chronic graft-vs-host disease activity and severity?

Aim To investigate whether increased YKL-40 levels positively correlate with graft-vs-host disease (cGVHD) activity and severity and if YKL-40 could serve as a disease biomarker.

Methods This case-control study was conducted at the University Hospital Centre Zagreb from July 2013 to October 2015. 56 patients treated with hematopoietic stem cell transplantation (HSCT) were included: 35 patients with cGVHD and 21 without cGVHD. There was no difference between groups in age, sex, median time from transplant to study enrollment, intensity of conditioning, type of donor, or source of stem cells. Blood samples were collected at study enrollment and YKL-40 levels were measured with ELISA. Disease activity was estimated using Clinician's Impression of Activity and Intensity of Immunosuppression scales and disease severity using Global National Institutes of Health (NIH) score.

Results YKL-40 levels were significantly higher in cGVHD patients than in controls ($P=0.003$). The difference remained significant when patients with myelofibrosis were excluded from the analysis ($P=0.017$). YKL-40 level significantly positively correlated with disease severity ($P<0.001$; correlation coefficient 0.455), and activity estimated using Clinician's Impression of Activity ($P=0.016$; correlation coefficient 0.412) but not using Intensity of Immunosuppression ($P=0.085$; correlation coefficient 0.296).

Conclusion YKL-40 could be considered a biomarker of cGVHD severity and activity. However, validation in a larger group of patients is warranted, as well as longitudinal testing of YKL-40 levels in patients at risk of developing cGVHD.

Nadira Duraković^{1,2},
Ivan Krečak³, Zinaida
Perić¹, Milan Milošević¹,
Lana Desnica², Dražen
Pulanić^{1,2,4}, Iskra Pusić⁵,
Vesna Kušec⁶, Radovan
Vrhovac^{1,2}, Steven Z.
Pavletic⁷, Damir Nemet^{1,2,4}

¹University of Zagreb, School of
Medicine, Zagreb, Croatia

²Department of Internal Medicine,
Division of Hematology, University
Hospital Center, Zagreb, Zagreb,
Croatia

³General Hospital Šibenik, Šibenik,
Croatia

⁴Faculty of Medicine Osijek, J. J.
Strossmayer University of Osijek,
Osijek, Croatia

⁵Division of Oncology, Section
of Bone Marrow Transplant
and Leukemia, Department of
Medicine, Washington University
School of Medicine, St Louis, MO,
USA

⁶Clinical Department of Laboratory
Diagnosis, University Hospital
Center Zagreb, Zagreb, Croatia

⁷Experimental Transplantation and
Immunology Branch, Center for
Cancer Research, National Cancer
Institute, National Institutes of
Health, Bethesda, MD, USA

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Correspondence to:

Nadira Duraković
University Hospital Centre Zagreb,
Dept of Internal Medicine, Division
of Hematology
Kišpatićeva 12
10000 Zagreb, Croatia
nadira@mef.hr

Chronic graft vs host disease (cGVHD) remains the most important cause of non-relapse morbidity and mortality in long-term survivors after hematopoietic stem cell transplantation (HSCT) (1,2) and by far its most intriguing complication. Although the precise immunologic mechanism leading to cGVHD development still remains to be elucidated, there have been some recent advances in understanding the disease process and identification of potential biomarkers (1-4). cGVHD is a multisystem disorder characterized by immune-dysregulation, resulting in impaired organ function, increased risk of infections, and deteriorated quality of life. Patients present with a variety of symptoms and organs involved including the skin, mouth, eye, gut, liver, lungs, joints, and genitourinary system (5-7). In the recent years the incidence of cGVHD has been increasing (2), likely related to the increased donor (8) and recipient age (9), decreased early post-transplant mortality, use of matched unrelated donors, and peripheral blood stem cell grafts (10,11). Identifying biomarkers that could be used to predict response to treatment, assess disease activity, or distinguish reversible disease activity from irreversible damage would be of great clinical value (12). Unfortunately, even though a number of potential biomarkers have been identified, such as anti-double-strand DNA antibodies, adiponectin, soluble IL-2 receptor α (IL-2R α), B-cell activating factor (BAFF), CXCL9, and CD13 (12-15), there is still no reliable marker that could be widely used in cGVHD patients.

Chitin, a polymer of N-acetylglucosamine, is present in coatings and cell walls of many organisms including bacteria, fungi, nematodes, insects, and plants (16-21). Chitinases, whose function is to degrade chitin, have been generally considered not to be present in mammals due to the absence of chitin. YKL-40, a member of the mammalian chitinase-like glycoproteins, is a heparin- and chitin-binding lectin without chitinase activity (22). It is expressed in various cell types including neutrophils (23), macrophages (24), bone marrow megakaryocytes (25), chondrocytes and synovial cells (26,27), as well as in malignant cells (28). In normal bone marrow, YKL-40 protein is stored in the granules of the myelocytes and metamyelocytes, and released from fully activated cells (29). YKL-40 is also expressed by macrophages *in vitro* during the late stage of differentiation (29), *in vivo* during inflammation (30), and by peritumoral macrophages (31). Furthermore, YKL-40 modulates vascular endothelial cell morphology by promoting the formation of branching tubules, acts as a chemoattractant for endothelial cells, stimulates their migration, and promotes the migration and adhesion of vascular smooth mus-

cle cells, indicating its role in angiogenesis (32). It has been shown to increase the growth rates of fibroblasts synergistically working with insulin-like growth factor-1 (IGF-1) (26). Its production is regulated by various cytokines. Studies in interleukin 6 (IL-6) knockout mice revealed that YKL-40 expression depended on IL-6 (33). Expression of YKL-40 mRNA in human monocyte is strongly stimulated by IFN γ , and inhibited by IL-4 and dexamethasone (34).

The physiological and biological functions of YKL-40 are still unclear. YKL-40 has been implicated in various inflammatory conditions, such as infections (35), autoimmune diseases (36-38), liver diseases (39,40) and malignant diseases (24,25,28,41-45). Mainly due to its role in inflammation (30) and extracellular matrix remodeling (26), it has been investigated as a potential biomarker of several autoimmune conditions (36,38), as well as those that include fibroblast activation (40,43).

In the non-myeloablative allogeneic HSCT setting, higher pretransplant recipient and donor plasma YKL-40 concentrations suggest a role for YKL-40 as a biomarker for relapse and treatment-related toxicity. Recipients with pretransplant YKL-40 concentrations above the age-adjusted 95th percentile (high) had higher relapse-related mortality and lower progression-free and overall survival. Recipients transplanted with donors with high YKL-40 concentrations had an increased probability and risk of grade 2-4 acute graft-vs-host disease (aGVHD) (45,46). However, none of the studies so far has examined whether post-transplant levels of YKL-40 influence the transplant outcomes or GVHD.

Based on the strong involvement of YKL-40 in inflammatory processes and autoimmune disorders, particularly given that YKL-40 production depends on IL-6 secretion and also IFN γ stimulation, we hypothesized that its expression was higher in patients with cGVHD than in transplanted patients without cGVHD and that it positively correlated with disease severity and activity.

PATIENTS AND METHODS

Patients

This case-control study is part of a larger project entitled "Clinical and Biological Factors Determining Severity and Activity of Chronic GVHD After Allogeneic Hematopoietic Stem Cell Transplantation" at the University Hospital Center Zagreb. The project included all patients who were referred to hematologist for post-transplantation follow up,

regardless of their age or underlying diagnosis, who consented to the study participation. Excluded from participation were patients with significant medical condition or any other significant circumstance that could affect the patient's ability to tolerate, comply, or complete the study and patients who according to the investigators assessment had life expectancy less than 3 months. Over the period of July 2013 to October 2015, 76 patients treated with hematopoietic stem cell transplantation (HSCT) were included in the project: 47 patients who developed cGVHD and 29 who did not develop cGVHD and who served as controls (Table 1).

For 56 patients (35 patients with cGVHD and 21 controls) included in the project serum samples were obtained at enrolment and stored. These patients were included in the study presented here. Prior to enrolment all participants signed the informed consent, and the study was approved by the University Hospital Center Zagreb Ethics Committee.

Data collection

Data regarding the diagnosis, time and type of transplant, and donor characteristics, and demographic data were collected. Blood samples for measurement of YKL-40 level and C-reactive protein (CRP) were taken at the time of study enrollment. For patients with established cGVHD diagnosis additional data regarding the severity and activity of disease were collected using predefined forms. Disease ac-

tivity was defined by Clinician's Impression of Activity and Intensity of Immunosuppression Scale. Clinician's impression of activity was defined as: inactive, off systemic therapy or topical immunosuppression; inactive, on systemic therapy or topical immunosuppression; active irrespective of the level of current therapy; and highly active irrespective of the level of current therapy (47). Intensity of immunosuppression scale was defined as: none; mild=single agent prednisone <0.5 mg/kg/d; moderate=prednisone \geq 0.5 mg/kg/d and/or any single agent/modality; high=2 or more agents/modalities \pm prednisone \geq 0.5 mg/kg/d (47,48). Disease severity was defined by Global National Institutes of health (NIH) scoring. Patients had mild cGVHD if only 1 or 2 organs (except lungs) were involved, with a maximum score 1 in all affected organs. Patients had moderate cGVHD if at least 1 organ was involved with clinically significant, but not major disability (maximum score 2) or 3 or more organs with no clinically significant functional impairment (maximum score 1 in all affected organs); a lung score 1 was classified as moderate. Patients had severe cGVHD if they had major impairment caused by cGVHD (score 3 in any organ); lung scores of 2 or 3 were classified as severe. Organs scored included the skin, eyes, mouth, gastrointestinal tract, liver, lungs, and joint/fascia. The genital area was scored only in women (49).

YKL-40 analysis

Plasma samples were prepared from EDTA (EDTA)-anticoagulated blood taken at the time of inclusion and were

TABLE 1. Characteristics of patients who underwent hematopoietic stem cell transplantation (HSCT) with and without chronic graft-vs-host disease (cGVHD)

		cGVHD (n=35)	Control (n=21)	P
Median age, years (range)		45 (9-60)	40 (16-59)	0.912
Sex	female, n (%)	18 (51.4)	10 (47.6)	0.783
	male, n (%)	17 (48.6)	11 (52.4)	
Diagnosis	aplastic anemia, n (%)	3 (8.6)	1 (4.8)	0.679
	acute lymphoblastic leukemia, acute myeloid leukemia, n (%)	21 (60.0)	10 (47.6)	
	chronic lymphocytic leukemia, n (%)	1 (2.9)	2 (9.5)	
	chronic myeloid leukemia and myeloproliferative diseases, n (%)	6 (17.1)	6 (28.6)	
	lymphoma, n (%)	2 (5.7)	2 (9.5)	
	immunodeficiencies, n (%)	1 (2.9)	0 (0.0)	
	multiple myeloma, n (%)	1 (2.9)	0 (0.0)	
Donor relationship	related, n (%)	21 (60.0)	9 (42.9)	0.213
	unrelated, n (%)	14 (40.0)	12 (57.1)	
Cell source	bone marrow, n (%)	15 (42.9)	8 (38.1)	0.726
	peripheral cells, n (%)	20 (57.1)	13 (61.9)	
Days from transplantation to enrollment, median (range)		463 (61-7853)	428 (190-1770)	0.441

stored at -80°C until YKL-40 analysis. YKL-40 plasma concentration was measured using a commercially available ELISA kit (R&D Systems Europe, Abingdon, UK).

Statistical analysis

After testing for normality using Kolmogorov-Smirnov test and due to small sample size we decided to use non-parametric tests. Categorical variables are presented as frequencies and corresponding percentages and quantitative variables as medians and interquartile ranges. The differences in categorical clinical parameters between patients and controls were analyzed using Fisher exact test or Fisher-Freeman-Halton exact test of independence when the contingency table was larger than 2×2 , while differences in quantitative variables were analyzed using Mann-Whitney U test. Differences in YKL-40 levels between NIH groups were analyzed using Kruskal-Wallis test. Spearman correlation coefficients were calculated to assess the correlation between YKL-40 levels and other clinical variables. *P* values below 0.05 were considered significant. Data analysis software system IBM SPSS Statistics, version 21.0 (IBM Corp., Armonk, NY, USA) was used.

RESULTS

Patient characteristics

Median age was 45 years (interquartile range 27-52 years) in the cGVHD group and 40 years (interquartile range 33-

54 years) in the control group. There were 18 women in cGVHD group and 9 in the control group. Graft stem cell source was the bone marrow in 15 (42.9%) and 8 (38.1%) and peripheral blood cells in 20 (57.1%) and 13 (61.9%) patients in cGVHD and control group, respectively. 17 (48.6%) and 8 (38.1%) patients received transplant from a related donor, while 18 (51.4%) and 13 (61.9%) patients received it from an unrelated donor in cGVHD and control group, respectively (Table 1). Myeloablative conditioning was used in 17 (48.6%) and 8 (38.1%) patients in cGVHD and control group, respectively. In the cGVHD group, 4 patients had mild, 15 moderate, and 16 severe symptoms as assessed by Global NIH score.

cGVHD and control patient groups were comparable according to age, sex, time from transplantation to enrollment, type of disease, cell source, donor relationship, intensity of conditioning, total body irradiation use in conditioning, and myelofibrosis as primary disease (Table 1).

Plasma YKL-40 concentration in cGVHD and control group

YKL-40 levels in cGVHD patients were significantly higher than in the control group (median 707 pg/mL [interquartile range 525-1054] vs 314 pg/mL [interquartile range 169-635]; $P=0.003$) (Figure 1A). Since YKL-40 is known to be increased in patients suffering from myelofibrosis, we excluded the data of all the patients with a history of myelofibrosis from the analysis (3 patients, [8.6%] in cGVHD

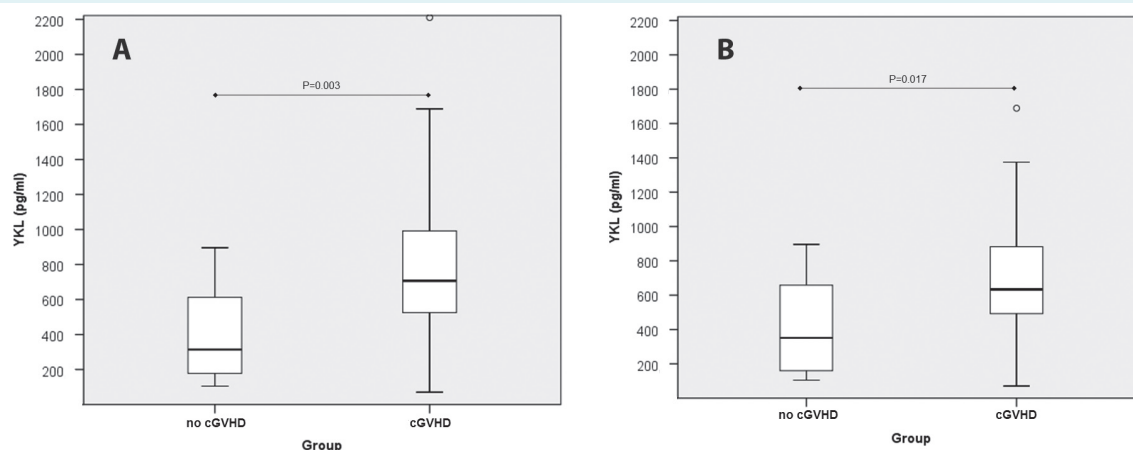


FIGURE 1. (A) Significantly higher YKL-40 serum concentration in chronic graft-vs-host disease (cGVHD) patients compared to control group. **(B)** The difference in YKL-40 serum concentration between cGVHD and control group remained significant after patients suffering from myelofibrosis were excluded from the analysis. 6 patients were excluded, 3 from each group.

group and 3 patients, [14.3%] in control group). After these patients were excluded, the concentration of YKL-40 was still significantly higher in cGVHD group than in the control group (median 633.5 pg/mL [range 71-1689] vs 351.5 pg/mL [range 105-2790]; $P=0.017$) (Figure 1B).

Correlation of YKL-40 concentration and disease activity and severity

YKL-40 levels significantly positively correlated with Clinician's Impression of Activity ($P=0.016$; correlation coefficient 0.412) and Global NIH score ($P<0.001$; correlation coefficient 0.455) (Figure 2), but not with Intensity of Immunosuppression ($P=0.085$; correlation coefficient 0.296). Since YKL-40 protein is involved in inflammation process, we tested its correlation with the CRP level, to establish if the YKL-40 level was increased due to persistent chronic inflammation not noticed by the examining physician and found no significant correlation ($P=0.581$) (Table 2).

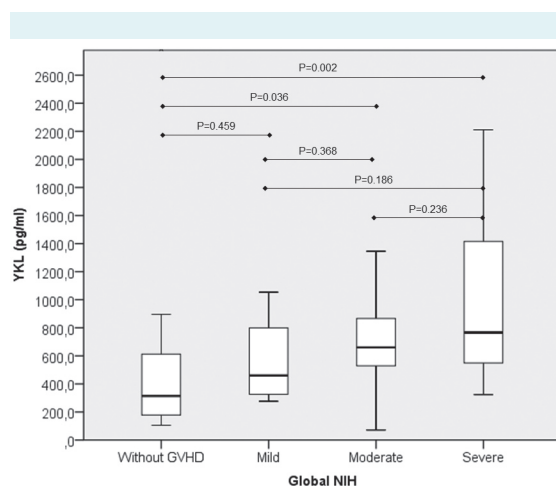


FIGURE 2. YKL-40 serum concentration was positively correlated with global National Institutes of Health score.

TABLE 2. Univariate analysis of YKL-40 serum concentration and chronic graft-vs-host disease (cGVHD) activity and severity measurements

	Correlation coefficient	P
Clinicians impression of activity	0.412	0.016
Intensity of immunosuppression	0.296	0.085
Global National Institutes of Health score	0.455	<0.001
National Institutes of Health average score	0.208	0.231
C reactive protein	0.097	0.581

DISCUSSION

Our study shows that the level of circulating YKL-40 is significantly higher in patients with cGVHD than in transplanted patients without cGVHD and correlates with disease severity and activity, as measured by Global NIH score and Clinicians Impression of Activity, respectively.

Although the exact role of glycoprotein YKL-40 in chronic inflammation is still not elucidated, YKL-40 concentration was found to be increased in 54% of patients with clinically active rheumatoid arthritis (RA). In patients in whom RA became inactive serum YKL-40 concentration decreased after 12 months, but increased in patients with RA flare (50). Furthermore, it has been shown that the level of circulating YKL-40 depends on IL-6 secretion, stimulated by IFN γ (45) and inhibited by IL-4 (33,34,51). Also, IL-6 and IFN γ have been shown to increase during GVHD development (52) and IL-6 is crucial for Th17 pathway (53), while production of IL-4 has been shown to be decreased in cGVHD patients (54). This could in part explain why YKL-40 is increased in active cGVHD reaction.

Furthermore, it was previously shown that patients with myelofibrosis had highly elevated levels of circulating YKL-40 in comparison to healthy controls (43). However, in our study the difference in plasma YKL-40 concentration between cGVHD group and control group remained significant after patients with myelofibrosis were excluded. The cause of elevated YKL-40 in myelofibrosis patients has not been found, but increased bone turnover and pronounced chronic inflammation have been implicated (43). Both processes are aborted after successful HSCT, and therefore, the fact that patient once had myelofibrosis should not influence the post-transplant level of YKL-40 protein. The effect of donor and recipient pre-transplant level of circulating YKL-40 on transplant outcomes (ie, relapse-related mortality, progression-free survival, overall survival, and acute GVHD incidence) has been reported, but post-transplant levels have not been investigated (45,46).

Even though our study distinctly shows concentration of YKL-40 to be significantly higher in patients with cGVHD in comparison to controls, it has several limitations. It is a single center study with a limited number of included patients. Albeit there was no significant difference between groups in time from transplant to sampling, there was a considerably wide range among patients within each group (range for cGVHD group was 61 to 7853 days, control 190 to 1770 days). This is because patients

were included at various points after having been diagnosed with cGVHD and at various stages of disease. Even though it would be preferable if the study included only newly diagnosed patients to limit the influence of disease advancement and/or therapy, the patients included in this study were clinically very well characterized according to 2005 NIH Consensus criteria.

In our study YKL-40 levels positively correlated with Global NIH score, a measure of disease severity, and Clinician's Impression of Activity, a measure of disease activity. We agree that data shown here do not prove YKL-40 protein to be a valid biomarker, but they certainly indicate that it is a strong new candidate. In our opinion, YKL-40 needs to be explored as a potential biomarker on a larger number of cGVHD patients. In addition, a longitudinal study of post-transplant patients, with serial, predefined time points is needed to validate these results, establish the dynamics of YKL-40 expression, and determine its role in evaluating the response to treatment.

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Ethical approval Received from the University Hospital Centre Zagreb and University of Zagreb School of Medicine.

Declaration of authorship ND designed the study, analyzed and interpreted the data, and wrote the manuscript. IK contributed through acquisition and analysis of data and drafting the manuscript. ZP contributed through data analysis and interpretation. MM contributed through data analysis and interpretation. LD contributed through data acquisition and interpretation, and drafting the manuscript. DP contributed through data acquisition and interpretation, and drafting the manuscript. IP contributed through data interpretation and drafting the manuscript. VK contributed through data acquisition and drafting the manuscript. RV contributed through conception and design of the study and drafting the manuscript. SZP contributed through data interpretation and drafting the manuscript. DN contributed through conception and design of the study and drafting the manuscript. All authors revised the work critically for important intellectual content, approved final version of the manuscript, and agreed to be accountable for all aspects of the work.

Competing interests All authors have completed the Unified Competing Interest form at www.icmje.org/coi_disclosure.pdf (available on request from the corresponding author) and declare: no support from any organization for the submitted work; no financial relationships with any organizations that might have an interest in the submitted work in the previous 3 years; no other relationships or activities that could appear to have influenced the submitted work.

References

- 1 Socié G, Stone JV, Wingard JR, Weisdorf D, Henslee-Downey PJ, Bredeson C, et al. Long-term survival and late deaths after allogeneic bone marrow transplantation. Late Effects Working Committee of the International Bone Marrow Transplant Registry. *N Engl J Med*. 1999;341:14-21. [Medline:10387937](#) [doi:10.1056/NEJM199907013410103](#)
- 2 Arai S, Arora M, Wang T, Spellman SR, He W, Couriel DR, et al. Increasing Incidence of chronic graft-versus-host disease in allogeneic transplantation: a report from the Center for International Blood and Marrow Transplant Research. *Biol Blood Marrow Transplant*. 2015;21:266-74. [Medline:25445023](#) [doi:10.1016/j.bbmt.2014.10.021](#)
- 3 Paczesny S, Hakim FT, Pidala J, Cooke KR, Lathrop J, Griffith LM, et al. National Institutes of Health Consensus Development Project on Criteria for Clinical Trials in Chronic Graft-versus-Host Disease: III. The 2014 Biomarker Working Group Report. *Biol Blood Marrow Transplant*. 2015;21:780-92. [Medline:25644957](#) [doi:10.1016/j.bbmt.2015.01.003](#)
- 4 Socié G, Ritz J. Current issues in chronic graft-versus-host disease. *Blood*. 2014;124:374-84. [Medline:24914139](#) [doi:10.1182/blood-2014-01-514752](#)
- 5 Atkinson K. Chronic graft-versus-host disease. *Bone Marrow Transplant*. 1990;5:69-82. [Medline:2178709](#)
- 6 Jagasia MH, Greinix HT, Arora M, Williams KM, Wolff D, Cowen EW. National Institutes of Health Consensus Development Project on Criteria for Clinical Trials in Chronic Graft-versus-Host Disease: I. The 2014 Diagnosis and Staging Working Group report. *Biol Blood Marrow Transplant*. 2015;21:389-401. [Medline:25529383](#) [doi:10.1016/j.bbmt.2014.12.001](#)
- 7 Jacobshon DA, Montross S, Anders V, Vogelsang GB. Clinical importance of confirming or excluding the diagnosis of chronic graft-versus-host disease. *Bone Marrow Transplant*. 2001;28:1047-51. [Medline:11781615](#) [doi:10.1038/sj.bmt.1703278](#)
- 8 Kollman C, Howe CW, Anasetti C, Antin JH, Davies SM, Filipovich AH, et al. Donor characteristics as risk factors in recipients after transplantation of bone marrow from unrelated donors: the effect of donor age. *Blood*. 2001;98:2043-51. [Medline:11567988](#) [doi:10.1182/blood.V98.7.2043](#)
- 9 Arora M, Klein JP, Weisdorf DJ, Hassebroek A, Flowers ME, Cutler CS, et al. Chronic GVHD risk score: a Center for International Blood and Marrow Transplant Research analysis. *Blood*. 2011;117:6714-20. [Medline:21493797](#) [doi:10.1182/blood-2010-12-323824](#)
- 10 Remberger M, Beelen DW, Fauser A, Basara N, Basu O, Ringdén O. Increased risk of extensive chronic graft-versus-host disease after allogeneic peripheral blood stem cell transplantation using unrelated donors. *Blood*. 2005;105:548-51. [Medline:15367429](#) [doi:10.1182/blood-2004-03-1000](#)
- 11 Ringdén O, Labopin M, Bacigalupo A, Arcese W, Schaefer UW, Willemze R, et al. Transplantation of peripheral blood stem cells as compared with bone marrow from HLA-identical siblings in adult patients with acute myeloid leukemia and acute lymphoblastic leukemia. *J Clin Oncol*. 2002;20:4655-64. [Medline:12488410](#) [doi:10.1200/JCO.2002.12.049](#)
- 12 Schultz KR, Miklos DB, Fowler D, Cooke K, Shizuru J, Zorn E, et al. Toward biomarkers for chronic graft-versus-host disease: National Institutes of Health consensus development project on criteria

- for clinical trials in chronic graft-versus-host disease: III. *Biol Blood Marrow Transplant*. 2006;12:126-37. [Medline:16443511](#) [doi:10.1016/j.bbmt.2005.11.010](#)
- 13 Fujii H, Cuvelier G, She K, Aslanian S, Shimizu H, Kariminia A, et al. Biomarkers in newly diagnosed pediatric-extensive chronic graft-versus-host disease: a report from the Children's Oncology Group. *Blood*. 2008;111:3276-85. [Medline:17925486](#) [doi:10.1182/blood-2007-08-106286](#)
 - 14 Nakasone H, Binh PN, Yamazaki R, Tanaka Y, Sakamoto K, Ashizawa M, et al. Association between serum high-molecular-weight adiponectin level and the severity of chronic graft-versus-host disease in allogeneic stem cell transplantation recipients. *Blood*. 2011;117:3469-72. [Medline:21258011](#) [doi:10.1182/blood-2010-10-316109](#)
 - 15 Kitko CL, Levine JE, Storer BE, Chai X, Fox DA, Braun TM, et al. Plasma CXCL9 elevations correlate with chronic GVHD diagnosis. *Blood*. 2014;123:786-93. [Medline:24363401](#) [doi:10.1182/blood-2013-08-520072](#)
 - 16 Broekaert WF, Van Parijs J, Leyns F, Joos H, Peumans WJ. A chitin-binding lectin from stinging nettle rhizomes with antifungal properties. *Science*. 1989;245:1100-2. [Medline:17838811](#) [doi:10.1126/science.245.4922.1100](#)
 - 17 Watanabe T, Suzuki K, Oyanagi W, Ohnishi K, Tanaka H. Gene cloning of chitinase A1 from *Bacillus circulans* WL-12 revealed its evolutionary relationship to *Serratia* chitinase and to the Type III homology units of fibronectin. *J Biol Chem*. 1990;265:15659-65. [Medline:2203782](#)
 - 18 Hayes CK, Klemsdal S, Lorito M, Di Pietro A, Peterbauer C, Nakas JP, et al. Isolation and sequencing of an endochitinase-encoding gene from a cDNA library of *Trichoderma harzianum*. *Gene (Amst)*. 1994;120:143-8. [doi:10.1016/0378-1119\(94\)90797-8](#)
 - 19 Kramer KJ, Corpuz L, Choi HK, Muthukrishnan S. Sequence of a cDNA and expression of the gene encoding epidermal and gut chitinases of *Manduca sexta*. *Insect Biochem Mol Biol*. 1993;23:691-701. [Medline:8353525](#) [doi:10.1016/0965-1748\(93\)90043-R](#)
 - 20 Krishnan A, Nair PN, Jones D. Isolation, cloning, and characterization of a new chitinase stored in the active form in chitin-lined venom reservoir. *J Biol Chem*. 1994;269:20971-6. [Medline:8063715](#)
 - 21 Jekel PA, Hartman JBH, Beintema JJ. The primary structure of heveamine, an enzyme with lysozyme/chitinase activity from *Hevea brasiliensis* latex. *Eur J Biochem*. 1991;200:123-30. [Medline:1879417](#) [doi:10.1111/j.1432-1033.1991.tb21057.x](#)
 - 22 Rehli M, Krause SW, Andreesen R. Molecular characterization of the gene for human cartilage gp-39 (CHI3L1), a member of the chitinase protein family and marker for late stages of macrophage differentiation. *Genomics*. 1997;43:221-5. [Medline:9244440](#) [doi:10.1006/geno.1997.4778](#)
 - 23 Volck B, Price PA, Johansen JS, Sorensen O, Benfield TL, Nielsen HJ. YKL-40, a mammalian member of the chitinase family, is a matrix protein of specific granules in human neutrophils. *Proc Assoc Am Physicians*. 1998;110:351-60. [Medline:9686683](#)
 - 24 Biggar RJ, Johansen JS, Smedby KE, Rostgaard K, Chang ET, Adami HO, et al. Serum YKL-40 and interleukin 6 levels in Hodgkin lymphoma. *Clin Cancer Res*. 2008;14:6974-8. [Medline:18980992](#) [doi:10.1158/1078-0432.CCR-08-1026](#)
 - 25 Mylin AK, Andersen NF, Johansen JS, Abildgaard N, Heickendorff L, Standal T, et al. Serum YKL-40 and bone marrow angiogenesis in multiple myeloma. *Int J Cancer*. 2009;124:1492-4. [Medline:19089918](#) [doi:10.1002/ijc.24110](#)
 - 26 Recklies AD, White C, Ling H. The chitinase 3-like protein human cartilage glycoprotein 39 (HC-gp39) stimulates proliferation of human connective tissue cells and activates both extracellular signal-regulated kinase - and protein kinase B-mediated signaling pathways. *Biochem J*. 2002;365:119-26. [Medline:12071845](#) [doi:10.1042/bj20020075](#)
 - 27 Volck B, Ostergaard K, Johansen, Garbarsch C, Price PA. The distribution of YKL-40 in osteoarthritic and normal human articular cartilage. *Scand J Rheumatol*. 1999;28:171-9. [Medline:10380840](#) [doi:10.1080/03009749950154257](#)
 - 28 Johansen JS, Schultz NA, Jensen BV. Plasma YKL-40: a potential new cancer biomarker? *Future Oncol*. 2009;5:1065-82. [Medline:19792974](#) [doi:10.2217/fon.09.66](#)
 - 29 Renkema GH, Boot RG, Au FL, Donker-Koopman WE, Strijland A, Muijsers AO. Chitotriosidase, a chitinase, and the 39-kDa human cartilage glycoprotein, a chitin-binding lectin, are homologues of family 18 glycosyl hydrolases secreted by human macrophages. *Eur J Biochem*. 1998;251:504-9. [Medline:9492324](#) [doi:10.1046/j.1432-1327.1998.2510504.x](#)
 - 30 Baeten D, Boots AMH, Steenbakkers PG, Elewaut D, Bos E, Verheijden GF, et al. Human cartilage gp-39+, CD16+ monocytes in peripheral blood and synovium. Correlation with joint destruction in rheumatoid arthritis. *Arthritis Rheum*. 2000;43:1233-43. [Medline:10857782](#) [doi:10.1002/1529-0131\(200006\)43:6<1233::AID-ANR6>3.0.CO;2-9](#)
 - 31 Junker N, Johansen JS, Andersen CB, Kristjansen PE. Expression of YKL-40 by peritumoral macrophages in human small cell lung cancer. *Lung Cancer*. 2005;48:223-31. [Medline:15829322](#) [doi:10.1016/j.lungcan.2004.11.011](#)
 - 32 Nishikawa KC, Millis AJT. gp38k (CHI3L1) is a novel adhesion and migration factor for vascular cells. *Exp Cell Res*. 2003;287:79-87. [Medline:12799184](#) [doi:10.1016/S0014-4827\(03\)00069-7](#)
 - 33 Kzyshkowska J, Mamidi S, Gratchev A, Kremmer E, Schmutzmaier C, Krusel L, et al. Novel stabilin-1 interacting chitinase-like protein (SI-CLP) is up-regulated in alternatively activated macrophages and secreted via lysosomal pathway. *Blood*. 2006b;107:3221-8. [Medline:16357325](#) [doi:10.1182/blood-2005-07-2843](#)
 - 34 Di Rosa M, Musumeci M, Scuto A, Musumeci S, Malaguarnera L. Effect of interferon-gamma, interleukin-10, lipopolysaccharide

- and tumor necrosis factor-alpha on chitotriosidase synthesis in human macrophages. *Clin Chem Lab Med*. 2005;43:499-502. [Medline:15899671](#) [doi:10.1515/CCLM.2005.088](#)
- 35 Ostergaard C, Johansen JS, Benfield T, Price PA, Lundgren JD. YKL-40 is elevated in cerebrospinal fluid from patients with purulent meningitis. *Clin Diagn Lab Immunol*. 2002;9:598-604. [Medline:11986266](#)
 - 36 Vind I, Johansen JS, Price PA, Munkholm P. Serum YKL-40, a potential new marker of disease activity in patients with inflammatory bowel disease. *Scand J Gastroenterol*. 2003;38:599-605. [Medline:12825867](#) [doi:10.1080/00365520310000537](#)
 - 37 Van Bilsen JH, van Dongen H, Lard LR, van der Voort EL, Elferink DG, Bakker AM, et al. Functional regulatory immune responses against human cartilage glycoprotein-39 in health vs. proinflammatory responses in rheumatoid arthritis. *Proc Natl Acad Sci U S A*. 2004;101:17180-5. [Medline:15569925](#) [doi:10.1073/pnas.0407704101](#)
 - 38 Grosso S, Margollicci MA, Bargagli E, Buccoliero QR, Perrone A, Galimberti D, et al. Serum levels of chitotriosidase as a marker of disease activity and clinical stage in sarcoidosis. *Scand J Clin Lab Invest*. 2004;64:57-62. [Medline:15025429](#) [doi:10.1080/00365510410004092](#)
 - 39 Malaguarnera L, Di Rosa M, Zambito AM, dell'Ombra N, Nicoletti F, Malaguarnera M. Chitotriosidase gene expression in Kupffer cells from patients with non-alcoholic fatty liver disease. *Gut*. 2006;55:1313-20. [Medline:16825325](#) [doi:10.1136/gut.2005.075697](#)
 - 40 Tran A, Benzaken S, Saint-Paul MC, Guzman-Granier E, Hastier P, Pradier C, et al. Chondrex (YKL-40), a potential new serum fibrosis marker in patients with alcoholic liver disease. *Eur J Gastroenterol Hepatol*. 2000;12:989-93. [Medline:11007134](#) [doi:10.1097/00042737-200012090-00004](#)
 - 41 Mylin AK, Andersen NF, Johansen JS, Abildgaard N, Heickendorff T, Standal T, et al. Serum YKL-40 and bone marrow angiogenesis in multiple myeloma. *Int J Cancer*. 2009;124:1492-4. [Medline:19089918](#) [doi:10.1002/ijc.24110](#)
 - 42 Bergmann OJ, Johansen JS, Klausen TW, Mylin AK, Kristensen JS, Kjeldsen E, et al. High serum concentration of YKL-40 is associated with short survival in patients with acute myeloid leukemia. *Clin Cancer Res*. 2005;11:8644-52. [Medline:16361549](#) [doi:10.1158/1078-0432.CCR-05-1317](#)
 - 43 Björn EM, Lykkesgaard Andersen C, Jensen MK, Hasselbalch HC. Circulating YKL-40 in myelofibrosis a potential novel biomarker of disease activity and the inflammatory state. *Eur J Haematol*. 2014;93:224-8. [Medline:24689875](#) [doi:10.1111/ejh.12332](#)
 - 44 Andersen CL, Björn ME, McMullin MF, Harrison C, Samuelsson J, Ejerblad E, et al. Circulating YKL-40 in patients with essential thrombocythemia and polycythemia vera treated with the novel histone deacetylase inhibitor vorinostat. *Leuk Res*. 2014;38:816-21. [Medline:24836761](#) [doi:10.1016/j.leukres.2014.04.002](#)
 - 45 Morup AM, Kornblit B, Johansen JS, Masmas TN, Madsen HO, Vindelov L, et al. The prognostic value of YKL-40 concentrations in nonmyeloablative conditioning allogeneic hematopoietic cell transplantation. *Biol Blood Marrow Transplant*. 2011;17:1299-307. [Medline:21232622](#) [doi:10.1016/j.bbmt.2011.01.008](#)
 - 46 Kornblit B, Tang W, Lee S. The prognostic value of YKL-40 in allogeneic hematopoietic cell transplantation. *Blood*. 2014;123.
 - 47 Grkovic L, Baird K, Steinberg SM, Williams KM, Pulanic D, Cowen EW, et al. Clinical laboratory markers of inflammation as determinants of chronic graft-versus-host disease activity and NIH global severity. *Leukemia*. 2012;26:633-43. [Medline:22005783](#) [doi:10.1038/leu.2011.254](#)
 - 48 Mitchell SA, Leidy NK, Mooney KH, Dudley WN, Beck SL, LaStayo PC, et al. Determinants of functional performance in long-term survivors of allogeneic hematopoietic stem cell transplantation with chronic graft-versus-host disease (cGVHD). *Bone Marrow Transplant*. 2010;45:762-9. [Medline:19784078](#) [doi:10.1038/bmt.2009.238](#)
 - 49 Filipovich AH, Weisdorf D, Pavletic S, Socie G, Wingard JR, Lee SJ, et al. National Institutes of Health consensus development project on criteria for clinical trials in chronic graft-versus-host disease: I. Diagnosis and staging working group report. *Biol Blood Marrow Transplant*. 2005;11:945-56. [Medline:16338616](#) [doi:10.1016/j.bbmt.2005.09.004](#)
 - 50 Johansen JS, Stoltenberg M, Hansen M, Florescu A, Horslev-Petersen K, Lorenzen I, et al. Serum YKL-40 concentration in patients with rheumatoid arthritis: relation to disease activity. *Rheumatology*. 1999;38:618-26. [Medline:10461474](#) [doi:10.1093/rheumatology/38.7.618](#)
 - 51 Henden AS, Hill GR. Cytokines in graft-versus-host disease. *J Immunol*. 2015;194:4604-12. [Medline:25934923](#) [doi:10.4049/jimmunol.1500117](#)
 - 52 Imamura M, Hashino S, Kobayashi H, Kubayashi S, Hirano S, Minagawa T, et al. Serum cytokine levels in bone marrow transplantation: synergistic interaction of interleukin-6, interferon-gamma, and tumor necrosis factor-alpha in graft-versus-host disease. *Bone Marrow Transplant*. 1994;13:745-51. [Medline:7920309](#)
 - 53 Chen X, Das R, Komorowski R, Beres A, Hessner MJ, Mihara M, et al. Blockade of interleukin-6 signaling augments regulatory T-cell reconstitution and attenuates the severity of graft-versus-host disease. *Blood*. 2009;114:891-900. [Medline:19491393](#) [doi:10.1182/blood-2009-01-197178](#)
 - 54 Rozmus J, Schultz KR, Wynne K, Karimnia A, Satyanarayanan P, Krailo M, et al. Early and late extensive chronic graft-versus-host disease in children is characterized by different Th1/Th2 cytokine profiles: findings of the Children's Oncology Group Study ASCT0031. *Biol Blood Marrow Transplant*. 2011;17:1804-13. [Medline:21669298](#) [doi:10.1016/j.bbmt.2011.05.011](#)