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Pulmonary lactate release following cardiopulmonary bypass

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ABSTRACT

Objective: The etiology of lung injury following cardiopulmonary bypass (CPB) is multifactorial. Our study focused on quantifying the lactate release from the lungs precipitated by extracorporeal circulation at different time points after the insult. This was complemented by an evaluation of the gas exchange at the level of the alveolar-capillary membrane.

Methods: Forty consecutive patients (age 61 ± 11 yrs, EUROScore 4.7 ± 2.7) undergoing CABG were prospectively analyzed. The data are presented as medians and the interquartile range.

Results: The pulmonary lactate release (PLR) increased from a baseline value of 0.033 [range 0.077 to 0.170] to 0.465 [range 0.113 to 0.922] mmol/min/m², which was seen 6 hours postoperatively ($P < 0.001$). The A-a O₂ gradient increased from 12.7 [range 8.8 to 15] to 39.1 kPa [range 30.3 to 46.5] upon discontinuation of CPB ($P < 0.001$). The systemic arterial lactate (L_s) concentration increased from 1.22 [range 1 to 1.44] to 3.03 [range 2.29 to 4.76] mmol/L six hours after surgery ($P < 0.001$). The venoarterial pCO₂ difference (V-A dpCO₂) rose from 0.6 [range 0.5 to 0.9] to 0.9 [range 0.7 to 1] kPa ($P = 0.014$). The mortality in the studied group was 5% (2/40).

Conclusions: The lungs were found to be a significant source of lactate, and this pulmonary lactate flux was accentuated by CPB. The PLR correlated with systemic hyperlactatemia as well as the A-a O₂ gradient, and was found to be higher in patients requiring prolonged mechanical ventilatory support. The duration of CPB had a significant impact on the systemic lactate concentrations, V-A dpCO₂, and the A-a O₂ gradient, but not on the PLR.

Key words: pulmonary lactate release, cardiopulmonary bypass, alveolar-arterial oxygen gradient

INTRODUCTION

Pulmonary dysfunction after cardiac surgery remains a frequent problem that may have a profound influence upon patient recovery. The wide spectrum of manifestations ranges from mild subclinical dysfunction to full-blown adult respiratory distress syndrome [1].

Multiple factors may precipitate acute lung injury following cardiopulmonary bypass, but the systemic inflammatory response is likely one of the most prominent causes [2]. Preoperative factors affecting the evolution of postoperative pulmonary dysfunction are chronic obstructive pulmonary disease, smoking and respiratory muscle wasting. Multiple features of the surgical procedure employing cardiopulmonary bypass harbor the potential to precipitate pulmonary injury. Hemodilution, reduction in the plasma oncotic pressure and left atrial pressure elevation associated with weaning from cardiopulmonary bypass (CPB) contribute to the accumulation of fluid in the lung interstitium [3,4]. The lungs are not immune to the effects of cytotoxic mediators, emboli or reperfusion injury induced by CPB. Furthermore, lung perfusion is altered in a way that it is solely dependant upon bronchial circulation, since the flow through the pulmonary artery is virtually nonexistent. The complex disruption of pulmonary homeostasis may be further enhanced by ventilator associated lung injury.

The pulmonary dysfunction may manifest itself by a widening of the alveolar-arterial oxygen gradient (A-a O₂ gradient), increase in pulmonary vascular resistance (PVR) or changes in the pulmonary shunt [2]. The most malignant form of CPB induced pulmonary dysfunction is the adult respiratory distress syndrome (ARDS). The incidence of ARDS following cardiac surgery is below 2% [1,5,6].

Serum lactate concentrations have been used as markers of systemic hypoperfusion in a variety of clinical settings, including the cardiac surgical patient population [7,8,9,10,11]. The etiology of hyperlactatemia following cardiopulmonary bypass, however, remains controversial. While the pathogenesis of type A lactic acidosis is unequivocally linked to hypoperfusion, type B lactic

acidosis, which occurs in patients in whom oxygen delivery is not impaired, may also occasionally be seen in patients undergoing CPB [12,13]. The lungs have previously been shown to be a source of lactate in the setting of lung injury, thereby producing a positive transpulmonary lactate gradient [14]. The venoarterial difference in the partial pressure of carbon dioxide (V-A dpCO_2) has been instrumental in demonstrating hypoperfusion in several published studies [15,16].

Our study aimed to test the hypothesis that cardiopulmonary bypass will have a significant impact on the pulmonary production of lactic acid, as well as compromise the functional integrity of the alveolar-capillary membrane. Furthermore, the pulmonary lactate release and the A-a oxygen gradient were documented at different time points in relation to the surgical insult in order to provide a chronological insight into the evolution of these markers of lung injury.

PATIENTS AND METHODS

Following the approval from our institutional ethics committee, 40 consecutive patients undergoing coronary artery bypass grafting using cardiopulmonary bypass (CABG) were enrolled into our study. Informed consent was obtained from all patients. The study was conducted in a prospective observational fashion. Exclusion criteria were concomitant valvular pathology requiring surgery and reoperative surgical myocardial revascularization.

Perioperative management

The patients received diazepam and morphine 30 minutes prior to induction of anesthesia. Endotracheal tube, urinary catheter, as well as radial artery and pulmonary artery catheters were inserted. The anesthetic regime included induction and maintenance of anesthesia with midazolam, fentanyl and pancuronium bromide. This was coupled with sevoflurane inhalation. The initial ventilator settings included a tidal volume of 8 ml/kg, and a respiratory rate of 12 breaths per minute. Typically, the FiO₂ was set at 50%. The critical components of the employed cardiopulmonary circuit were the Medtronic Affinity Trillium membrane oxygenator, venous reservoir and PVC tubing (Medtronic, Minneapolis, USA) and a Stoeckert III roller pump (Stoeckert, Munich, Germany). Systemic heparinization aiming at an ACT > 480 seconds was used, followed by full reversal with protamine after decannulation. Tepid CPB was employed, targeting the flow at 2.2 L/min/m². The target mean arterial pressure during CPB was 60 mmHg. If necessary, norepinephrine was employed to reach the aimed blood pressure. The surgical myocardial revascularization was performed on an arrested heart, during a period of aortic cross-clamping. The lungs were open to atmosphere during CPB. Weaning from CPB was initiated once the patient's rhythm had stabilized and normothermia had been achieved. Inotropic support was initiated in order to maintain a cardiac index greater than 2.2 L/min/m². The preferred inotropic agent was dobutamine. Norepinephrine was used if dobutamine produced excessive

vasodilatation. Epinephrine was used if the hemodynamic performance remained inadequate with the previously mentioned catecholamines. An intraaortic ballon pump was inserted if further support was required.

Measurement and Calculations

The study protocol consisted of obtaining arterial blood gas samples from the radial artery and mixed venous blood samples from the distal port of the pulmonary artery catheter.

Thermodilution cardiac indices were obtained in triplicate and then averaged. The blood samples from the radial and pulmonary arteries were drawn simultaneously. The first sampling of radial and pulmonary arterial blood was performed upon induction of anesthesia and prior to the surgical incision (T1). Values obtained from the preoperative sampling and the results thus obtained, were used as control values. The second sample (T2) was taken intraoperatively, upon discontinuation of cardiopulmonary bypass and resumption of intrinsic cardiac and pulmonary function. The third (T3) and fourth (T4) samples were drawn in the intensive care unit 6 and 18 hours after surgery, respectively. Thermodilution cardiac indices were obtained at all of the above mentioned stages of blood sampling. The raw data was then processed to obtain the transpulmonary lactate gradient and the pulmonary lactate release from the following formulas:

$$\text{TPLG} = \text{Lactate}_{\text{RA}} - \text{Lactate}_{\text{PA}} - ((\text{Hb}_{\text{RA}} - \text{Hb}_{\text{PA}}) / \text{Hb}_{\text{RA}}) \times \text{Lactate}_{\text{RA}}$$
$$\text{PLR} = \text{TPLG} \times \text{CI}$$

CI = cardiac index

Hb_{RA} = radial artery hemoglobin

Lactate_{PA} = lactate concentration in the pulmonary artery

Lactate_{RA} = lactate concentration in the radial artery

Hb_{PA} = pulmonary artery hemoglobin

PLR= Pulmonary lactate release

TPLG= Transpulmonary lactate gradient

The effects of hemodilution were taken into account in order to obtain a hemoglobin adjusted pulmonary lactate release as was suggested by Kellum et al [14]. The A-a oxygen gradient was

obtained by subtracting the partial pressure of oxygen in arterial blood from the calculated alveolar oxygen tension [17]. The formulas for calculating the A-a oxygen gradient are shown here:

$$\begin{aligned} \text{A-a O}_2 \text{ gradient} &= P_{\text{A}}\text{O}_2 - P_{\text{a}}\text{O}_2 \\ P_{\text{A}}\text{O}_2 &= P_{\text{i}}\text{O}_2 - P_{\text{A}}\text{CO}_2/R \\ P_{\text{i}}\text{O}_2 &= (P_{\text{B}} - P_{\text{H}_2\text{O}}) \times \text{FiO}_2 \end{aligned}$$

A-a O₂ gradient = Alveolar arterial oxygen gradient
P_AO₂ = Alveolar oxygen tension
P_aO₂ = Partial pressure of oxygen in arterial blood
P_iO₂ = Partial pressure of inspired oxygen
P_ACO₂ = Alveolar carbon dioxide tension (assumed to equal the partial pressure of CO₂ in

arterial blood due to the ease of exchange of CO₂)
R = Respiratory quotient (assumed at 0.8)
P_B = Barometric pressure
P_{H₂O} = Water pressure (6.2 kPa as inspired air is fully saturated at the level of the carina)
FiO₂ = Fractional concentration of inspired oxygen

One sample of arterial blood was analyzed at any given time point to obtain the A-a oxygen gradient. Individual A-a oxygen gradients were recorded for every patient at every time point. The data from all of the patients were then pooled and analyzed in order to obtain median values for the whole cohort at every preset time point (T1-T4). The initial FiO₂ was 0.5 for the samples taken prior to the surgical incision. The subsequent fraction of inspired oxygen was dictated by each patient's present partial pressure of oxygen in arterial blood. This value of FiO₂ was then incorporated into the calculation for the corresponding A-a oxygen gradient. Only one FiO₂ value was used for one time point, i.e. no attempt to investigate the effects of different ventilation settings on the A-a oxygen gradient was made.

The venoarterial pCO₂ difference was obtained by subtracting the partial pressure of CO₂ in radial arterial blood from the partial CO₂ pressure measured in the pulmonary artery.

Statistical analysis

The data are presented as mean values ± standard deviation or medians followed by the interquartile range for variables with a distribution that is not normal. Longitudinal comparisons

between samples of the same subject were analyzed using the ANOVA test. Correlations were analyzed using the nonparametric Spearman R correlation methodology. Analyses between different groups of patients were performed using the Mann-Whitney U test. A $P < 0.05$ was considered to be of statistical significance. The data were processed using the Statistica software package (StatSoft Inc., Tulsa, USA).

RESULTS

Perioperative summary

Forty consecutive patients undergoing coronary artery bypass grafting (CABG) were enrolled into our study. The patient demographic data are presented in Table 1. The spectrum of comorbidities seen in our patient population is typical for the contemporary cardiac surgical practice.

The patient population profile included both elective and non-elective cases. There were four patients in whom the operation was performed on an emergent basis, related to complications of percutaneous coronary interventions. All of these had acute coronary syndromes noted prior to surgery. Two of these patients were found to be in significant hemodynamic compromise when referred to our tertiary care center from outside hospitals. Two patients had intraaortic balloon pumps placed preoperatively. The mortality was 5% (2/40). One of these patients died due to left ventricular free wall rupture on postoperative day 14, whereas the second death in this cohort was stroke related. The perioperative summary is presented in Table 2.

The median values of the pulmonary lactate release increased from a baseline value of 0.033 [range -0.077 to 0.170] to 0.465 [range 0.113 to 0.922] observed at six hours after surgery ($P < 0.001$). The transpulmonary lactate gradient (TPLG) increased from the preoperative 0.014 [range -0.034 to 0.085] to 0.163 [range 0.041 to 0.249] mmol/L calculated from the samples taken 6 hours after surgery ($P = 0.001$).

The A-a O₂ gradient increased from a baseline value of 12.7 [range 8.8 to 15] to 39.1 kPa [range 30.3 to 46.5], immediately upon discontinuation of cardiopulmonary bypass ($P < 0.001$). The total arterial lactate content measured in the radial artery increased from 1.22 [range 1 to 1.44] to 3.03 [range 2.29 to 4.76] mmol/L at 6 hours postoperatively ($P < 0.001$). The serum lactate concentration did not reach preoperative values by the end of the study protocol. A rise in the

venoarterial pCO₂ gradient (V-A dpCO₂) from 0.6 [range 0.5 to 0.9] to 0.9 [range 0.7 to 1] kPa at 6 hours postoperatively was also found to be statistically significant ($P = 0.014$). This peak corresponded to the simultaneous peak in the systemic lactate concentration. There was no statistical significance to the dynamics of the pulmonary vascular resistance throughout the studied period. The evolution of the pulmonary lactate flux, transpulmonary lactate gradient, A-a O₂ gradient, total arterial lactate, venoarterial pCO₂ difference and PVR from the values obtained prior to CPB to those found at 18 hours postoperatively are presented in more detail in Table 3.

Clinical correlations

The amplification of the pulmonary lactate release correlated with the simultaneously observed increase in the systemic arterial lactate concentration at 6 (T3) hours after surgery. The PLR calculated from samples immediately upon discontinuation of CPB (T2) correlated with the appropriate A-a O₂ gradient. The peak PLR also correlated with the A-a O₂ gradient observed at 18 hours after surgery. The PLR^{T4} correlated with the venoarterial pCO₂ difference, which served as another index of systemic hypoperfusion. A lower A-a O₂ gradient prior to surgery was predictive of quicker extubation. The alveolar-arterial oxygen gradient seen at 18 hours correlated with the duration of cardiopulmonary bypass. The A-a oxygen gradient^{T4} correlated with the corresponding pH^{T4} in addition to having an inverse correlation with the partial pressure of carbon dioxide in arterial blood sampled at the same time (T4). The peak increase in the systemic arterial lactate concentration (T3) correlated with the duration of CPB as well as the duration of mechanical ventilation. Additionally, the duration of CPB influenced the PVR and V-A dpCO₂ at 18 hours after CABG. The correlations between the above noted measurements and clinical parameters are presented in greater detail in Table 4.

The subgroup of patients in whom mechanical ventilation for more than 72 hours was required had a higher pulmonary lactate release 6 hours after surgery in comparison to patients weaned from the ventilator before that time (5.15 [range 0.991 to 5.956] vs. 0.392 [range 0.096 to 0.766])

mmol/min/m², Mann-Whitney U test $P = 0.005$). Additionally, this subgroup of patients had higher A-a O₂ gradients at 18 hours after surgery (32.5 [range 24.1 to 33.5] vs. 17.6 [range 11.3 to 20.5], $P < 0.001$), higher serum lactate concentrations at T3 and T4 (13.63 [range 8.36 to 15.4] vs. 2.85 [range 2.27 to 4.04] mmol/L, $P < 0.001$, and 7.42 [range 4.26 to 8.2] vs. 1.97 [range 1.3 to 2.71] mmol/L, $P < 0.001$, respectively) as well as higher pulmonary vascular resistance 6 hours after surgery (172 [range 121 to 200] vs. 92 [range 65 to 123] dyne·sec/cm⁵, $P = 0.048$). Higher EUROScore and CPB times were also noted in patients requiring prolonged ventilatory support (9±1 vs. 4.4±2.5, $P = 0.008$, and 105±37 vs. 66±26 min, $P = 0.035$). Patients in whom an intraaortic balloon pump was inserted had higher peak lactate levels (8.73 [range 5.28 to 15.4] vs. 2.85 [range 2.27 to 4.04], $P = 0.005$).

DISCUSSION

The genesis of respiratory complications following cardiac surgery is complex and may stem from inherent pulmonary disease, depressed cardiac function with resultant pulmonary edema, post-perfusion inflammatory response or inadequate diaphragmatic function associated with phrenic nerve injury [2,5]. The functional and clinical changes have been corroborated by histological evidence of lung injury consisting of alveolar edema, extravasation of red blood cells and neutrophils as well as necrosis and swelling of pneumocytes and endothelial cells [1].

Cardiopulmonary bypass is responsible for the production and release of a plethora of markers, which serve to quantify both local organ dysfunctions and systemic perfusion aberrancies.

Our study focused on the lactate release from the lungs and the dysfunction at the level of the alveolar-capillary membrane precipitated by extracorporeal circulation in patients undergoing surgical myocardial revascularization. In addition, we looked at systemic hypoperfusion indices, such as systemic lactate concentration and veno-arterial pCO₂ gradient, and their correlation to cardiopulmonary bypass.

We have demonstrated a modest pulmonary lactate production in our patient population even at baseline, i.e. the lungs were a source of lactate even in the absence of a notable surgical insult.

The pulmonary lactate contribution rose significantly upon exposure to surgical trauma and cardiopulmonary bypass. The occurrence of a positive transpulmonary lactate gradient after CPB has been corroborated by other authors [14,18].

The values continued to rise up to six hours after the procedure, implying that the lactate washout from the lungs may be a protracted process. The clinical impact of the pulmonary lactate release was illustrated in its correlation with the need for prolonged mechanical ventilatory support.

There was no correlation between the PLR and the duration of CPB. While the lactate produced in the lungs was not the sole factor responsible for systemic hyperlactatemia, it significantly contributed to the systemic arterial lactate content measured at six after the procedure. The

dramatic augmentation of the alveolar-arterial oxygen gradient after surgery may corroborate the hypothesis that cardiopulmonary bypass caused a complex disturbance at the level of the alveolar-capillary membrane. One must recognize, however, that the presence of atelectasis after resumption of ventilation may have also contributed to this finding. Persistent widening of the alveolar-arterial O₂ gradient was indicative of the requirement for prolonged mechanical ventilation. We were able to show a statistically significant correlation between the A-a O₂ gradient and the PLR after weaning from CPB. Arterial hyperlactatemia reflects the undesirable effects of CPB at the systemic level. We have demonstrated a correlation of the peak systemic lactate concentration with the duration of CPB and mechanical ventilation. Not surprisingly, patients in whom an intraaortic balloon pump for mechanical circulatory support was inserted had higher peak lactate levels. One should not neglect the possible confounding effect of catecholamine administration on the observed hyperlactatemia. Surprisingly, however, we found no statistically significant correlation between the use of inotropic support and systemic lactate concentrations. The evidence of global hypoperfusion during CPB in our study also included widening of the difference between the mixed venous and arterial pCO₂ difference. The V-A dpCO₂ correlated with the duration of CPB.

We did not maintain pulmonary artery (PA) perfusion during cardiopulmonary bypass in our patients, nor were the lungs ventilated. Maintaining physiologic pulmonary perfusion has been advocated by some authors, in order to preserve the pulmonary clearance of certain proinflammatory cytokines [1]. The PA perfusion strategy has been linked to superior postoperative respiratory function in certain clinical scenarios, possibly due to attenuation of the ischemia-reperfusion injury, but has yet to gain wider acceptance [19,20]. The higher mortality and morbidity seen in our cohort is secondary to the patient profile referred to our center which accepts patients suffering from various complications from smaller regional hospitals, and is also reflective of the prospective nature of this study. Four patients in this cohort were referred for

surgery suffering from various degrees of myocardial ischemia and hemodynamic compromise due to either failed PCI in the setting of an acute coronary syndrome or complications thereof. There are certain limitations in this study that warrant mentioning. The pulmonary lactate release was calculated from the lactate gradient between the afferent and efferent circulations in relation to the lungs. The efferent sample, however, was not taken from the pulmonary veins, but rather from the radial artery. Theoretical concern could therefore be raised whether shunting through Thebesian veins could compromise the validity of the data. This issue, however, has been addressed in the past and the mixed venous sample has been utilized for quantification of the pulmonary lactate production in several studies [14,18,21]. Additionally, the sample size is relatively small; with the subgroup of patients requiring prolonged ventilatory support smaller still.

In summary, our study is the first to offer a chronological perspective on the pulmonary lactate release after cardiopulmonary bypass in a relatively homogenous population of patients. The main findings of this study are that the pulmonary lactate release is accentuated by the surgical procedure employing cardiopulmonary bypass and that a higher PLR is predictive of prolonged mechanical ventilation. It also showed a correlation between the PLR and the disturbance at the level of the alveolar-capillary membrane, as well as the impact of pulmonary lactate production on systemic hyperlactatemia. While there is no doubt that cardiopulmonary bypass induces a violent disturbance of the normal cardiovascular physiology, one should not rush to condemn it as the sole etiologic factor responsible for the development of pulmonary dysfunction. One must also stress that the amplitude of pulmonary lactate production in our study is much less pronounced than that observed in patients with profound respiratory insufficiency documented in another study encompassing a wide range of etiologies [14].

Table 1. Preoperative patient characteristics

Age (yrs)	61 ± 11
Gender (n/%)	
Male	34 (85)
Female	6 (15)
EUROScore	4.7 ± 2.7
Ejection fraction	53 ± 10
Hypelipidemia (n/%)	18 (45)
Hypertension (n/%)	28 (70)
Diabetes mellitus (n/%)	10 (25)
Smoking history (n/%)	16 (40)
Preop. AMI (n/%)	21 (53)
Emergent surgery	4 (10)
Left main stenosis (n/%)	11 (28)
Three vessel disease (n/%)	32 (80)
Mild-moderate MR (n/%)	17 (43)

Table 2. Perioperative patient data presented as mean values \pm standard deviation, unless otherwise indicated

No. of grafts	3 \pm 0.8
Cross clamp time (minutes)	37 \pm 12
CPB time (minutes)	69 \pm 28
Inotropic support (n/%)	18 (45)
ICU (days)*	2 (2/3)
Mechanical ventilation (hours)*	8 (7/14)
Complications (n/%)	
Stroke	1 (3)
Periop. myocardial ischemia	5 (13)
Reexploration for bleeding	2 (5)
Acute renal failure	2 (5)
Sternal wound infection	1 (3)
Prolonged mechanical ventilation (>72 hrs)	3 (8)
IABP requirement	3 (8)
Death (n/%)	2 (5)

*Data as median and quartiles

Table 3. Longitudinal evolution of studied variables

Variables	T1	T2	T3	T4
PLR <i>mmol/min/m²</i>	0.033 [-0.077/0.170]	0.335 [-0.285/0.973]	0.465 [0.113/0.922]	0.421 [-0.032/0.945]
TPLG <i>mmol/L</i>	0.014 [-0.034/0.085]	0.105 [-0.102/0.279]	0.163 [0.041/0.249]	0.120 [-0.01/0.298]
A-a O ₂ G <i>kPa</i>	12.7 [8.8/15]	39.1 [30.3/46.5]	18.2 [12.1/21.6]	17.7 [11.7/21.7]
Systemic lactate <i>mmol/L</i>	1.22 [1/1.44]	2.54 [2.06/3.26]	3.03 [2.29/4.76]	2.07 [1.34/2.93]
V-A dpCO ₂ <i>kPa</i>	0.6 [0.5/0.9]	0.6 [0.4/0.8]	0.9 [0.7/1]	0.7 [0.5/1]
PVR <i>dyne·sec/cm⁵</i>	114 [72/148]	94 [70/124]	98 [66/125]	95 [71/125]

Data are shown as median with quartiles

T1= Preoperative value
T2= Value after discontinuation of CPB
T3= Value 6 hrs postop
T4= Value 18 hrs postop

PLR = Pulmonary lactate release
TPLG= Transpulmonary lactate gradient
A-a O₂G= Alveolar-arterial oxygen gradient
V-A dpCO₂= Venoarterial pCO₂ difference
PVR = Pulmonary vascular resistance

Table 4. Clinical relevance of laboratory data tested with a nonparametric correlation analysis.

Only correlations with a $P < 0.05$ are listed.

Variables	Spearman R Correlation	<i>P</i>
PLR ^{T2} & A-a G O ₂ ^{T2}	0.39	0.012
PLR ^{T3} & A-a G O ₂ ^{T4}	0.48	0.002
PLR ^{T3} & Ls ^{T3}	0.43	0.006
PLR ^{T4} & A-a G O ₂ ^{T4}	0.32	0.047
PLR ^{T4} & V-A dpCO ₂ ^{T4}	0.37	0.019
TPLG ^{T2} & A-a G O ₂ ^{T2}	0.42	0.007
TPLG ^{T3} & A-a G O ₂ ^{T4}	0.52	0.001
TPLG ^{T3} & Ls ^{T3}	0.43	0.006
TPLG ^{T4} & A-a G O ₂ ^{T4}	0.33	0.035
TPLG ^{T4} & Ls ^{T4}	0.40	0.011
TPLG ^{T4} & V-A dpCO ₂ ^{T4}	0.38	0.015
A-a O ₂ G ^{T1} & M.vent	-0.46	0.003
A-a O ₂ G ^{T4} & CPB	0.34	0.029
A-a O ₂ G ^{T4} & PaCO ₂ ^{T4}	-0.33	0.039
A-a O ₂ G ^{T4} & pH ^{T4}	0.33	0.035
Ls ^{T2} & CPB	0.35	0.027
Ls ^{T3} & CPB	0.45	0.004
Ls ^{T3} & M.vent	0.44	0.004
Ls ^{T4} & M.vent	0.51	0.001
Ls ^{T4} & PVR ^{T4}	0.38	0.018
PVR ^{T4} & CPB	0.35	0.027
V-A dpCO ₂ ^{T4} & CPB	0.44	0.005

T1= Preoperative value
T2= Value after discontinuation of CPB
T3= Value 6 hrs postop
T4= Value 18 hrs postop

PLR = Pulmonary lactate release
PaCO₂ = Partial pressure of carbon dioxide
A-a O₂ G = Alveolar-arterial oxygen gradient
L_s = Systemic arterial lactate
TPLG= Transpulmonary lactate gradient
V-A dpCO₂ = Veno-arterial pCO₂ difference
M.vent = Duration of mechanical ventilation
PVR = Pulmonary vascular resistance

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